REVIEW

New insights into Chlorella vulgaris applications

Mohammed Al-Hammadi^{[1](http://orcid.org/0009-0009-4317-3431)} | Mine Güngörmüşler² \bullet

1 Division of Bioengineering, Graduate School, Izmir University of Economics, Izmir, Türkiye

²Department of Genetics and Bioengineering, Faculty of Engineering, Izmir University of Economics, Izmir, Türkiye

Correspondence

Mine Güngörmüşler, Department of Genetics and Bioengineering, Faculty of Engineering, Izmir University of Economics, Sakarya Caddesi No. 156, Izmir 35330, Türkiye. Email: mine.gungormusler@ieu.edu.tr

Abstract

Environmental pollution is a big challenge that has been faced by humans in contemporary life. In this context, fossil fuel, cement production, and plastic waste pose a direct threat to the environment and biodiversity. One of the prominent solutions is the use of renewable sources, and different organisms to valorize wastes into green energy and bioplastics such as polylactic acid. Chlorella vulgaris, a microalgae, is a promising candidate to resolve these issues due to its ease of cultivation, fast growth, carbon dioxide uptake, and oxygen production during its growth on wastewater along with biofuels, and other productions. Thus, in this article, we focused on the potential of Chlorella vulgaris to be used in wastewater treatment, biohydrogen, biocement, biopolymer, food additives, and preservation, biodiesel which is seen to be the most promising for industrial scale, and related biorefineries with the most recent applications with a brief review of Chlorella and polylactic acid market size to realize the technical/nontechnical reasons behind the cost and obstacles that hinder the industrial production for the mentioned applications. We believe that our findings are important for those who are interested in scientific/financial research about microalgae.

KEYWORDS

biocement, biodiesel, biofuel, bioplastic, biorefinery, wastewater treatment

1 | INTRODUCTION

Fossil fuels have a significant impact on human health (Rajagopalan & Landrigan, [2021](#page-14-0)) and environmental issues, leading to nearly one in five deaths worldwide and 8.7 million premature deaths annually (Chaisson, [2021\)](#page-11-0). Fuels and cooking technology pose serious health risks to 3 billion people, leading to 7 million fatalities in 2016. Heart disease, cancer, stroke, chronic illnesses, and respiratory problems are the leading causes (Sekar et al., [2021\)](#page-15-0). Food production accounts for 15% of fossil fuel use (Shagun, [2023\)](#page-15-1), with agriculture and food handling accounting for 21% and 49% of total US food production, respectively (Save On Energy, [2022](#page-15-2)). The pollution generated by fuels reduces photosynthesis, hindering crop growth and causing

major staple crops to lose 110 million tons annually, 4% of global crop production, and potentially 15% in some regions (CCAC, [2023\)](#page-11-1). Cement and plastic production are other sources that contribute to air pollution. More than 4 billion tons of cement are produced annually, contributing to about 8% of global $CO₂$ emissions (Chatham House, [2018\)](#page-11-2) and its production is growing by 2.5% each year (Rubenstein, [2012](#page-14-1)). While 400 million tons of plastic are produced annually (IUCN, [2021](#page-12-0)), almost 12 million barrels of oil are consumed annually to produce plastic bags (Roberts, [2020](#page-14-2)). Over 99% of plastics are fossil fuel-based (CHO, [2020\)](#page-11-3), with 14 million tons ending up in the ocean annually. Humans have produced 8.3 billion tons of plastic since the 1950s, with 6.3 billion tons discarded. Only 9% has been recycled and 12% incinerated, increasing production from 2

This is an open access article under the terms of the [Creative Commons Attribution](http://creativecommons.org/licenses/by/4.0/) License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2024 The Authors. Biotechnology and Bioengineering published by Wiley Periodicals LLC.

million tons in the 1950s to 4 million tons in 2015. 79% of plastic waste is found in the natural environment, with an estimated 12 billion tons by 2050 (Mullarkey, [2017](#page-14-3)). Polymers, including fossil and bio‐based plastics, are classified as biodegradable if over 60%–70% degrade within 6 months (Awaja et al., [2004](#page-10-0)). Bio‐based polymers can be produced from plants (Ehman & Area, [2021](#page-11-4)), and animals (Machineni & Rao Anupoju, [2022\)](#page-13-0). Despite the fact that not all bio‐ based plastics are biodegradable,. Approximately half of the current bio-based plastics on the market are not (Rahman & Bhoi, [2021](#page-14-4)). Microbial fermentation is a promising method for producing bio‐ based, degradable polymers, such as polylactic acid (PLA), by converting corn into dextrose through wet milling and fermentation (Gotro, [2012](#page-12-1)). PLA is now a commercial bioplastic made of 100% biological materials, making it suitable for mixing with microalgae (Bulota & Budtova, [2015\)](#page-11-5). Polyhydroxyalkanoates (PHAs) biodegradable biopolymer can also be obtained from oiled and de‐ oiled microalgae biomass; further, microalgae can be employed in biorefineries to reduce the cost of the processes (Chew et al., [2017](#page-11-6)), making it a sustainable technology for companies and investors since they aim for long‐term profitability, continuous improvements, and market recognition (Pavolová et al., [2021\)](#page-14-5). Technological shocks that are described as abrupt technological advancements that have a major impact on social, political, economic, or other outcomes play a direct role in investment as well (Akbari et al., [2021\)](#page-10-1). However, the literature lacks information on recent advancements in using Chlorella vulgaris, microalgae known for its ease of cultivation and fast‐growing features (Ma'mun et al., [2022\)](#page-13-1), for biofuels, food additives correlated with health, biocement, and biopolymer production, covering technical and some financial obstacles that hinder Chlorella vulgaris from being utilized on industrial scales, addressing a crucial question regarding the most practical industrial application technically and economically and the future research needed to make certain applications more practical to resolve the present issues with plastic and fossil fuel usage.

2 | CHLORELLA VULGARIS

C. vulgaris is a green eukaryotic microalga of the genus Chlorella that has existed on Earth since the Precambrian period (Safi et al., [2014\)](#page-15-3). These algae were found by Martinus Willem Beijerink in 1890 as the first microalgae with a well‐defined nucleus (Beyerinick, [1890](#page-10-2)). C. vulgaris requires macro- and micronutrients to grow. The macronutrients involve nitrogen (N), carbon (C), and phosphorus (P) (Aguda et al., [2023\)](#page-10-3), while the micronutrients involve inorganic trace elements (Hong et al., [2016\)](#page-12-2). Moreover, C. vulgaris requires a tolerated range of temperature, light intensity, and pH with optimum values of 25-30°C (Ma et al., [2014](#page-13-2)) and 6.5–9 (Jiang et al., [2021](#page-12-3)), respectively. It has the ability to grow under 2500–1000 lux of light intensity, including a 16:8 light:dark cycle (Febrieni et al., [2020\)](#page-11-7). Microalgae cultivation faces challenges such as availability of nutrients, temperature, light source, water availability, harvesting, and cost. Open‐pond cultivation is not

AL‐HAMMADI and GÜNGÖRMÜŞLER | 1487 **BIOENGINEERIN**

> preferred over lab-scale due to the microalgal contamination risk. On the other hand, a 1‐hectare lab‐scale cultivation is required due to the high costs of operation, process, and maintenance (Maroušek et al., [2023](#page-13-3)). Photobioreactors (PBRs) are transparent systems that allow producers to control production conditions by separating them from the external environment. They offer higher yield without contamination risk and cost about 1.4 million USD per hectare. However, their increased building costs make them unlikely to offer a competitive advantage in lower‐priced nations (Maroušek et al., [2023](#page-13-4)). Producing 1 ton of dry algae biomass was estimated to be 500 and 110 USD for the PBRs and open pond, respectively, considering the price of $CO₂$ capture when integrated into biomass production, which was 250 and 55 USD for the photobioreactor and open pond, respectively (Zabochnicka et al., [2022\)](#page-16-0).

> C. vulgaris is a spherical microscopic cell with a 5‐ to 10‐μm diameter (Scragg et al., [2003\)](#page-15-4) and has many elements similar to plants (Dvoretsky, Akulinin, et al., [2016\)](#page-11-8). Its cell wall is crucial for maintaining cell integrity and providing protection against invaders and harsh environments (Safi et al., [2014\)](#page-15-3). Moreover, the cell wall properties affect microalgae applications such as biodiesel production. Biodiesel can be derived from triglycerides by its conversion to biodiesel in the trans‐esterification process (John Love & Bryant, [2017\)](#page-12-4), and C. vulgaris is an attractive model for triglyceride studies due to its high triglyceride accumulation (Unterlander et al., [2017\)](#page-15-5). However, its complex cell wall significantly obstructs lipid extraction (Dvoretsky, Dvoretsky, et al., [2016](#page-11-9)). Furthermore, cell wall composition is also important in bioplastic production (Rahman & Miller, [2017\)](#page-14-6). Its 51-58% protein content makes it more crackresistant and thermally stable than Spirulina which is utilized to blend the biomass for bioplastic formation (Zeller et al., [2013\)](#page-16-1). Therefore, there were some efforts to control the protein content in C. vulgaris by increasing the ratio of the light intensity using red and white LED lamps (Metsoviti et al., [2020\)](#page-13-5). Following the cell wall, a gel-like substance known as cytoplasm, which is composed of water, soluble proteins, and minerals that contain microalgae organelles, namely, mitochondria, chloroplast, Golgi body, vacuoles, and a nucleus, exists (Safi et al., [2014](#page-15-3)). Mitochondria and chloroplasts are crucial in microalgae growth, regulating respiration and carbon dioxide uptake, which impacts lipid, biomass, and bioactive compound productivity. Mitochondria have their own DNA for respiration (Lewin & Andersen, [2022](#page-13-6)), while chloroplasts are responsible for photosynthesis and have their own DNA as well (Safi et al., [2014\)](#page-15-3). C. vulgaris lacks evidence of adapting to produce photosynthetic pigments to absorb light intensities (Rendón et al., [2013\)](#page-14-7). Genetic engineering tools have shown promising results in improving photosynthesis, with a 1.2‐fold increase in capacity and growth (Yang et al., [2017](#page-16-2)). Further study is needed to understand the function of mitochondria and chloroplasts under nuclear genome control and their own genomes for short and long-term effects.

> In the early 1990s, German scientists considered C. vulgaris as a new food source. Japan is currently the largest consumer of Chlorella sp. for food (Safi et al., [2014](#page-15-3)) and medicinal purposes (Freitas, [2017\)](#page-12-5).

TABLE 1 Major contributors in the Chlorella sp. market worldwide (Meticulous Research, [2022](#page-13-8)).

Company	Country
Flora Manufacturing & Distributing Ltd	Canada
Roquette Klötze GmbH & Co. KG	Germany
Phycom	Netherlands
Tianjin Norland Biotech Co., Ltd.	China
Fuging King Dnarmsa Spirulina Co. Ltd.	China
Far East Microalgae Industries, Co., Ltd. (FEMICO)	Taiwan
Allmicroalgae-Natural Products, S.A	Portugal
Qingdao Haizhijiao Biotechnology Co., Ltd.	China
STAUBER	U.S.
Alver World SA	Switzerland
AlgoSource	France
Taiwan Chlorella Manufacturing Company	Taiwan
E.I.D.-Parry	India
Dongtai City Spirulina Bio-engineering Co., Ltd.	China
Duplaco B.V.	Netherlands
Sun Chlorella Corporation	Japan
Algorigin	Switzerland
Yunnan Green A Biological Project Co., Ltd	China
Zhejiang Comp Spirulina Co., Ltd.	China
Gong Bih Enterprise Co., Ltd.	China
Wilson Group	Taiwan

Notably, the Chlorella sp. market started to be attractive recently. The size of the global chlorella sp. market was estimated at USD 275.21 million in 2021 and is anticipated to increase to USD 506.99 million by 2030, rising at a CAGR of 6.3% over the forecast period (2023–2030) from USD 292.55 million in 2022. Chlorella sp. application for food and beverages industry occupied the largest share in 2021, followed by personal care industry, nutraceuticals and Pharmaceuticals, and others, respectively. The demand for Chlorella sp. applications in medicine and personal care, is expected to increase market size in Northern America and Europe, respectively, and currently, Europe occupies 24.3% of Chlorella sp. market globally. In addition to that, several promising companies have stated about biodiesel production from algae which makes micro‐macro algae highlighted candidates for the studies and the potential of their utilization in the fuel sector. ExxonMobil and Solix Biofuels in the USA stated about producing 1500 gallons and 5000–8000 gallons yearly, respectively (Abdo et al., [2022;](#page-10-4) ExxonMobil, [2018](#page-11-10)). Euglena Co., Ltd in Japan stated about producing 76,650 gallons (from waste cooking oil and algae) yearly (Euglena, [2018\)](#page-11-11). The major contributors in the Chlorella sp. market worldwide are mentioned in Table [1](#page-2-0) below (Skyquest, [2022\)](#page-15-6).

FIGURE 1 Harvesting techniques for microalgae.

3 | HARVESTING TECHNIQUES

Harvesting is an important factor in utilizing microalgae on the industrial scale because of the cost (Fasaei et al., [2018](#page-11-12)), risk of contamination (Wan et al., [2015](#page-16-3)), and effect on the final product (Hidayah Mat Yasin et al., [2019](#page-12-6)). Harvesting expenses make up about 90% of the cost of the equipment used in biomass production, and they make up about 30% of the total cost of producing microalgae (Ma et al., [2023\)](#page-13-7), while harvesting of microalgae oil makes up about 60% of the produced oil (biodiesel) (Dewayanto et al., [2023\)](#page-11-13). Different techniques can be used for harvesting microalgae as shown in Figure [1](#page-2-1) below (Barros et al., [2015](#page-10-5)). Flocculation is a cost-effective, simple, and efficient method for large-scale biomass harvesting (Chozhavendhan et al., [2022\)](#page-11-14). Positively charged flocculants can destabilize and neutralize the negatively charged surface of microalgae, forming flocs for harvesting (Pugazhendhi et al., [2019\)](#page-14-8). Flocculation can be categorized into physical, chemical, and biological flocculation, depending on the harvested material (Branyikova et al., [2018](#page-10-6)). Chitosan, a commonly used organic polymer in C. vulgaris harvesting (Rashid et al., [2013](#page-14-9)), is an expensive flocculant and impractical for large‐scale production due to its high cost (Yin et al., [2020](#page-16-4)). It costs 20–50 USD per kg and 7280 USD to harvest a ton of microalgae; however, it can be optimized for coating, functionalizing micro and nanospheres, and dual harvesting with additional flocculants like clay. Changing chitosan into nano‐chitosan can reduce costs to \$24.6 per ton, making biodiesel production economically feasible (de Morais et al., [2023\)](#page-14-10). Cationic starch is also a synthetic organic polymer that can be used to harvest C. vulgaris (Huang et al., [2019](#page-12-7)). The acidic environment weakens electrostatic forces between microalgae and flocculates, making the process pH‐ dependent. The harvesting efficiency of C. vulgaris using cationic starch reached 99% at pH 11, with 25.74 g L⁻¹ flocs concentration. The maximum biomass flocculation capacity was 8.62 g at pH 3 utilizing 1g of starch, with self-flocculation. pH also affected the diameter of C. vulgaris flocs, with 0.553 mm achieved at pH 11, larger than 0.208 mm at pH 3 (Huang et al., [2019](#page-12-7)). A study suggests that the use of electrolytic microbubbles can enhance the efficiency of harvesting cationic starch for C. vulgaris, which is 5.5 times larger than the flocculation‐settling process but requires more energy (Wei

et al., [2020](#page-16-5)). Although cationic starch grafted tannin and nano‐ chitosan were documented to be very cheap, cost 27.4 and 24.6 USD per ton for microalgae harvesting, no reports of their commercial use exist (de Morais et al., [2023](#page-14-10)). Polymeric compounds can be economically feasible flocculants except forpolymeric flocculants if the ionic strength of culture suspension is high (Mathimani & Mallick, [2018](#page-13-9)). Therefore, inorganic polymers such as polyelectrolyte EM1, poly aluminum chloride, and polyacrylamide were proposed for this purpose, even though they have a risk of pollution and pH dependency (Wan et al., [2015\)](#page-16-3). Metal salts like ferric sulfate, aluminum sulfate, and ferric chloride are non‐polymeric chemicals used in microalgae flocculation. Aluminum sulfate is cost-effective, costs 28 USD per ton (de Morais et al., [2023\)](#page-14-10), but requires high doses, potentially leading to biomass contamination with aluminum or iron (Ummalyma et al., [2016\)](#page-15-7). Physical flocculation using ultrasound, electro‐flocculation, and magnetic nanoparticles was also suggested to overcome the chemical drawbacks; nevertheless, some have disadvantages as well. Electro-flocculation is unsuitable for industrial scales, since electro‐flocculation requires energy consumption (Fayad et al., [2017\)](#page-11-15), while ultrasound is hard to apply for large scales (Bosma et al., [2003](#page-10-7)). Magnetic nanoparticles are one of the best ways to harvest microalgae due to their possibility in dustrial‐ scale applications and eco-friendly (Wang et al., [2015\)](#page-16-6). Yet, there are recent efforts to lower the cost and energy consumption of magnetic nanoparticle synthesis and minimize the utilization of reagents during the harvesting process to make it more practical for the industrial scale. It was documented that the lowest cost is 347 USD per ton by magnetic nanoparticle fabrication and modification using arginine (de Morais et al., [2023\)](#page-14-10). Coated and naked magnetic nanoparticles were compared for C. vulgaris harvesting, detachment, and recycling of the nanoparticles. Yttrium iron oxide (Y₃Fe₅O₁₂) was more efficient in terms of harvesting efficiency; it reached 90%, while naked iron oxide $(Fe₃O₄)$ was much easier to recycle under higher pH values (Zhu et al., [2019\)](#page-16-7). Diethylaminoethyl (DEAE) and polyethyleneimine (PEI) showed a high harvesting efficiency as well (>90%). While the detachment was possible only for DEAE beads (Prochazkova et al., 2013). Recent evidence stated that $Fe₃O₄$ nanoparticles coated with PEI had a high efficiency which was up to 99%. However, the study did not include detachment or recycling efficiency (Gerulová et al., 2022). Furthermore, microwave-synthesized naked Fe₃O₄ particles were evaluated. The particles were prepared using a rapid and low-cost method, precipitating iron sulfate using NaOH and drying in the microwave. The harvesting efficiency was over 99% at pH 3.0, and the particles could be recycled and reused at least five times. However, the detachment process involves chloroform addition (Savvidou et al., [2021\)](#page-15-8), which should be replaced or avoided. Therefore, new investigations into the detachment process are required. Bio‐flocculation is another way to harvest C. vulgaris by different techniques such as auto‐flocculation by controlling pH by adding alkali or controlling the consumption of $CO₂$ to increase the pH which leads to the sedimentation and aggregations of microalgae (Mathimani & Mallick, [2018\)](#page-13-9), although, requires alkaline addition, and involves magnesium precipitation. It is unrelated to the changes in

AL-HAMMADI and GÜNGÖRMÜŞLER **AL-HAMMADI and GÜNGÖRMÜŞLER BIOENGINEERIN**

10970290, 2024, 5, Downloaded from https

wiley om/doi/10

.1002/bit.2866 by Izmir Ekor

, Wiley

Online Library

109/0204.3. Downloads the program with the control control control control of the control cont

Terms and Conditions (https

//onlinelibrary.wiley

sand-c onditions)

on Wiley Online Library

for rules of use; OP articles ing.

 $\frac{1}{5}$

applicable Creativ

on [17/04/2024]. See the

microalgal cells, and the term "auto‐flocculation" was suggested to be misleading (Vandamme et al., [2012](#page-15-9)). Microorganisms can be used as bio‐flocculants as well. The harvesting efficiency was 99% and 100% by utilizing Aspergillus fumigatus and Aspergillus sp. UMN F01, respectively (Chen et al., [2018](#page-11-16)). However, the mechanism of bioflocculation using microorganisms has not been fully defined; it is thought to be mainly a function of the reactivity of the extracellular biopolymer and/or the direct adsorption of the self-aggregating microorganisms on the target algae (Wan et al., [2015](#page-16-3)). Moreover, the high cost of bioflocculation which is 1350 USD per ton, adds another issue that should be addressed in the future and minimized to make it more practical for the scaling up.

4 | APPLICATIONS

4.1 | Bioremediation integration with biodiesel production

Bioremediation is a subfield of biotechnology that uses live organisms like bacteria and microorganisms to remove contaminants, pollutants, and toxins from soil, water, and other environments (Brown, [2022\)](#page-11-17). Algae are well‐known as autotrophs (organisms that utilize the energy from photosynthesis to grow), play a significant role in wastewater treatment (Amaro et al., [2023\)](#page-10-8). Algae's chemical treatment capabilities through photosynthesis, can fix carbon dioxide and efficiently remove excess nutrients at a low cost. Moreover, they produce oxygen, which can mitigate the biological oxygen demand (BOD) of wastewater. Algae use waste as food and break down contaminants through enzymes, uptaking nutrients like nitrogen and phosphorus (Sahu, [2014\)](#page-15-10) to grow and produce biomass, which can be utilized for different applications, including biodiesel (Anand et al., [2023](#page-10-9)). The choice of microalgae for wastewater treatment depends on the tolerance of a specific species to wastewater and its ability to grow and absorb nutrients from wastewater (Ariyanti et al., [2012\)](#page-10-10). For instance, Chlamydomonas mexicana, Scenedesmus obliquus, Chlamydomonas pitschmannii, and C. vulgaris were cultivated with diazinon, and it was found that C. vulgaris showed better growth with a higher removal of diazinon by 94% (Kurade et al., [2016\)](#page-13-10). The bioavailability of nutrients like phosphorus in wastewater is crucial for preventing phosphorus production, reducing costs. Plus, the concerning phosphorus depletion on Earth has brought attention to the bioavailability of phosphorus. Phosphorus can be available in different forms, minerals and organic. However, not all organic and mineral sources are accessible to micro‐macro organisms, which must be addressed for sustainable technology (Stávková & Maroušek, [2021\)](#page-15-11). C. vulgaris has been tested recently in integrated systems for different types of wastewater treatment with lipids investigation and biodiesel production simultaneously. When C. vulgaris was grown in textile wastewater (TWW), the results showed the highest growth in 50% diluted TWW reaching 11.07 mg g^{-1} of fatty acids methyl ester; on the other hand, C. vulgaris could remove methylene blue from undiluted TWW up to 99%, chemical oxygen

demand (76%), phosphorous, and nitrogen more than 80%, producing 9.12 mg g^{-1} of fatty acids methyl ester which included palmitic acid C16:0 and linolenoic acid C18:3. Thus, the authors asserted that utilizing undiluted wastewater is more practical (Fazal et al., [2021\)](#page-11-18).

The potential of dairy wastewater was investigated for lipid accumulation and fatty acids characterizations for biodiesel production; the authors stated that the maximum accumulation of lipids was seen in effluent with 50% dilution for initial and secondary concentration (93%, 86%) correspondingly in concentrations of 13 and 26 million cells per mL of algae, while the highest percentage of nitrate removal was 57% (Khalaji et al., [2021\)](#page-12-9), and before this study, the fatty acids methyl ester in the lipids were analyzed for biodiesel production when C. vulgaris was cultivated and compared with C. sorokiniana pa.91 in non‐sterilized effluent of dairy wastewater with light intensity optimization. The maximum and initial lipid content for C. sorokiniana pa.91 and C. vulgaris was 35%, 37%, and 31%, 34% at 2500 lux, respectively, and according to the amount of cetane number, the authors showed that the lipid qualities are effective for biodiesel and the fuel has a great potential even at low temperatures according to cold flow plugging properties (CFPP), and cloud point (CP) values (Asadi et al., [2020\)](#page-10-11). However, the literature lacks some information about a practical extraction for lipids and biodiesel production, along with productivity and yield amounts, by utilizing dairy wastewater. Municipal and urban wastewater are not less important than dairy wastewater because they can also provide the microalgae with the required nutrients. Municipal wastewater was employed for water treatment along with $CO₂$ capture and biodiesel production in a coupled system to reduce the cost of the process. The maximum growth was achieved at a C/N ratio of 4 with 5% (v/v) $CO₂$ concentration, nitrate removal of 81%, ammonium removal of more than 99%, and phosphate removal of 88%. The obtained fatty acid methyl esters number was C16 to C18 showing the potential for biodiesel production (Ayatollahi et al., [2021\)](#page-10-12). However, urban wastewater was indicated to its potential for biodiesel production when C. vulgaris was cultivated and the lipids formed 8% of the utilized dry biomass (3.7 g) along with nutrients removed by 87% of phosphate, 99% of ammoniacal nitrogen, and nitrate, and the authors stated that biomass is suitable to be utilized as a feedstock for biodiesel production considering the amount of the obtained fatty acid methyl esters (Ariana, [2016](#page-10-13)). Nevertheless, new investigations are required further to realize the optimization effect on the properties of the produced biodiesel and the characterization of fatty acid methyl esters because the literature lacks these data. Pilot‐ scale (open raceway ponds) was also investigated for swine wastewater treatment along with carbon dioxide capture for biodiesel production. At 3% CO₂, C. vulgaris MBFJNU-1 produced more microalgal biomass (478.5 mg L⁻¹) and total fatty acids (21%), had higher CO2 bio-fixation productivity (63.2 mg L⁻¹ d⁻¹) and lipid production (9.1 mg L $^{-1}$ d $^{-1}$), and nutrients removal (total phosphorus, 28%; total nitrogen, 82%; chemical oxygen demand, 37%). Additionally, enzymatic transesterification of wet biomass with 5% lipase TL and 5% phospholipase PLA resulted in the maximum biodiesel conversion (93%) (Xie et al., [2022\)](#page-16-8). However, the study did not assess

the biochemical makeup of microalgae cultivated in swine wastewater, a crucial factor for wastewater treatment and high-value product production simultaneously. Additionally, a border investigation is needed for pilot-scale ponds, as most studies focus on labscale comparisons. Scientific research is crucial for producing and utilizing biofuels at a low cost. However, the competitiveness of biofuels is also influenced by the price of fossil fuels. High fossil fuel prices make biofuels more economically viable, while a decrease may reduce their marketability. Government policies and investment can also impact the biofuel market, as increased investment in biofuel technologies can lead to higher oil prices (Vochozka, Horák, et al., [2020\)](#page-15-12). The EUR/USD exchange rate is also a significant factor, as it is heavily reliant on global oil prices, which affects all markets, as many nations use these currencies as their reserve currency (Vochozka et al., [2020\)](#page-15-13).

4.2 | Biohydrogen

Hydrogenase is an enzyme produced by green algae that plays an important role in anaerobic metabolism which can catalyze the reversible oxidation of molecular hydrogen (promotes the formation and utilization of hydrogen) (Wittkamp et al., [2018\)](#page-16-9). This enzyme is produced by green microalgae by coupling photosynthetic electron transport chains and plastid hydrogenase [Fe‐Fe] to generate hydrogen gas which differs from other enzymes produced by other microorganisms such as [Ni‐Fe] hydrogenase, and nitrogenase. Hydrogenase [Fe‐Fe] is more active than [Ni‐Fe] hydrogenase and nitrogenase by 10−100 times and by 1000 times, respectively (Li et al., [2022\)](#page-13-11), due to the existence of a distinctive active center (cluster H) (Khetkorn et al., [2017\)](#page-12-10). Consequently, biohydrogen production was investigated in microalgae. C. vulgaris can be utilized to produce biohydrogen in many ways (direct and indirect) due to the ability of microalgae to produce hydrogenase. In a direct way, biohydrogen can be directly produced during photosynthesis from the water. However, the accumulation of the simultaneously produced oxygen inhibits hydrogenase enzymes. In an indirect way, the biomass of the microalgae can be utilized by other microorganisms by dark fermentation to produce hydrogen. The major drawback of this method is the biomass pretreatment requirement which can be done by different methods (Figure [2](#page-5-0) below) because the hydrolytic enzymatic activity of hydrogen‐producing bacteria is typically low, the pretreatment step is frequently necessary for the hydrolysis of algal biomass to release the organic materials from the algal cells and make them readily biodegradable to be utilized further in the fermentation. Thus, it is considered one of the challenges that should be handled in biohydrogen studies (Wang & Yin, [2018\)](#page-16-10). Nevertheless, biorefinery employment can be more practical to reduce the cost of this process. For instance, the biomass of C. vulgaris, Scenedesmus obliquus, and Consortium C was converted into a carbon source for a bacterial strain, namely Enterobacter aerogenes by fermenting the microalgal biomass after cultivating the microalgae in urban wastewater for treatment. The sugar was accumulating after

FIGURE 2 Pretreatment methods for algal biomass for biohydrogen production via the fermentation process.

reaching the status of nutritional stress in the bioreactor and afterward, the fermentation took place, and the highest yield of biohydrogen was 56.8 mL $H₂/gVS$ by Scenedesmus obliquus followed by Consortium C and C. vulgaris with 46.8 mL H₂/gVS and 40.8 mL H2/gVS, respectively, while the highest‐accumulated biohydrogen production potential was for Scenedesmus obliquus (2.96 mL) followed by Consortium C and C. vulgaris with (2.91 mL) and (2.35 mL), respectively. Interestingly, C. vulgaris biomass reached a high production rate (2.9 mL h⁻¹), but due to the longer lag phase which was observed compared to other strains, the accumulated biohydrogen was lower than theirs as it was already mentioned (Batista et al., [2015](#page-10-14)). Therefore, more investigations on different composition media with different dilutions to figure out the productivity of biohydrogen of C. vulgaris biomass should be investigated since the culture media can affect the overall carbohydrates concentration in the biomass and eventually affect the fermentation outcome. Additionally, the pretreatment procedure for microalgae biomass might have a substantial impact on the microalgae's composition and biohydrogen output since it was investigated for Scenedesmus obliquus, and Chlorella sorokiniana which was uncovered for C. vulgaris (Wang & Yin, [2018](#page-16-10)). Another strategy to produce biohydrogen by C. vulgaris is sulfur‐deprived cells because sulfur deficiency is a major factor in the cultures of microalgae that plays a role in biohydrogen production since it can inhibit protein synthesis and causes severe stress effects, leading to degradation of the photosynthetic apparatus (Nagy et al., [2018\)](#page-14-12) which is known as complicated machinery that contains several complexes of protein–pigment that is involved in photosynthesis process (Rochaix, [2016\)](#page-14-13), and eventually reduces the production of oxygen, which is an inhibitor of the hydrogenase enzyme (Antal & Lindblad, [2005](#page-10-15)). When immobilized‐sulfur‐deprived cultures of C. vulgaris were cultivated to produce biohydrogen, the maximum rate was 34.8 mL L⁻¹ h⁻¹ (Rashid et al., [2011\)](#page-14-14). However, when artificial wastewater was utilized for immobilized-sulfurdeprived cultures of C. vulgaris, the production rate was notably decreased to 1.63 mL L⁻¹ h⁻¹; nevertheless, further investigation is

AL-HAMMADI and GÜNGÖRMÜŞLER **in termişi birinde**lerinin birinde birinde birinde birinde birinde birinde birinde b BIOENGINEERIN

109/0204.3. Downloads the program with the control control control control of the control cont 10970290, 2024, 5, Downloaded from https: nlinelibrary.wiley.com/doi/10.1002/bit.28666 by Izmir Ekonomi Univ Wiley Online Library on $[17.04/2024]$. See the Terms and Conditions (https //onlinelibrary.wiley and-conditions) on Wiley Online Library for rules of use; OA

articles ing. governed by è

applicable Creativ

required because the growth conditions such as light: dark cycle, light type, and intensity were not similar (Ruiz‐Marin et al., [2020\)](#page-14-15). Recently, the potential of biohydrogen production by C. vulgaris without sulfur deficiency was also highlighted using direct photosynthesis without sulfur deficiency in the media or exposing the cells to stress conditions, and biohydrogen was obtained in the dark and light conditions average of 2.08 and 4.98 mL L⁻¹ h⁻¹ for the dark and light conditions, respectively. While the maximum rate reached 12 mL L⁻¹ h⁻¹ (Touloupakis et al., [2021\)](#page-15-14). Even though hydrogen mobility is now more affordable costing 7 Euro per 100 km than traditional fossil fuels 15.6 Euro per 100 km for the first time in the European Union, hydrogen is still in its early stage in the majority of nations which impedes the advancement of hydrogen infrastructure worldwide (Maroušek et al., [2022\)](#page-13-12). The low yield and high cost of biohydrogen production limit its industrial scale production and make it uncompetitive with other biofuels like biodiesel besides, the challenges including biohydrogen purification, storage, delivery, and safety, must be addressed to ensure hydrogen's viability and competitiveness (Feng et al., [2023\)](#page-12-11). It has been demonstrated that nanoparticles enhance biohydrogen synthesis in microbial systems by acting as catalysts, promoting enzymatic processes that produce hydrogen (Maroušek, [2022\)](#page-13-13), and enhancing the biomass productivity of the microalgae (Hidalgo et al., [2023](#page-12-12)). However, certain concentrations can be toxic. Therefore, more investigations related to optimization are required.

4.3 | C. vulgaris supplements and extracts for food preserving

C. vulgaris is versatile food source rich in functional macro‐micro nutrients, including protein, fatty acids, and minerals (Panahi et al., [2016\)](#page-14-16) and is rich in vitamins (Salvia et al., [2014\)](#page-15-15) with more folate than spinach (Bito et al., [2020](#page-10-16)). The anti-lipid effects (lowering bad cholesterol and triglyceride) of C. vulgaris were investigated in many studies (Sherafati et al., [2022](#page-15-16)). In this regard, C. vulgaris powder was examined in mice, and it was observed that increases in serum and liver total cholesterol and triglycerides were significantly suppressed in the given C. vulgaris mice when were fed by a highfat diet of powdered C. vulgaris, and there was not a significant difference in high-density lipoprotein (HDL) with no significant effect in the endogenous metabolism (Chovančíková & Šimek, [2001](#page-11-19)). On the other hand, it was indicated that the addition of C. vulgaris to atorvastatin treatment (reducing the body's capacity to produce cholesterol by decreasing this production) for 8 weeks of the therapy did not associate with improved control of serum lipid profiles (Panahi et al., [2012\)](#page-14-17). Once, the anticancer properties of C. vulgaris extracts were highlighted by increasing the levels of the proteins Bax, P53, and caspase‐3 while decreasing the amount of the protein Bcl‐2, which causes apoptosis and DNA damage (Yusof et al., [2010\)](#page-16-11), and for this reason, the function of C. vulgaris extracts as antioxidant and anticancer were also investigated on HeLa and DPPH cell lines. The results showed that some extracts of C. vulgaris exhibited more than

50% anticancer activity and antioxidant activity against Hela cancer cell lines (El‐fayoumy et al., [2021](#page-11-20)). Furthermore, water extracts from Spirulina platensis and C. vulgaris showed prolonged characteristics of packaged sardine fish when they were investigated due to the availability of mineral content and bioactive chemical components. The extracts prolonged the shelf life of sardine for 3 days. However, the utilization of water extracts from S. platensis was preferred over the extracts of C. vulgaris. Nevertheless, both extracts can be used as antimicrobial and antioxidant additives to preserve sardine fish because these extracts inhibit the growth of lactic acid bacteria, while C. vulgaris extract was a stronger inhibitory on tyramine accumulation, with 2‐ to 4‐fold lower tyramine accumulation in sardine meat than S. platensis extract. Also, the authors indicated that C. vulgaris extract was more effective in the inhibition of ammonia production and the most toxic biogenic amines such as tyramine and histamine in fish meat (Özogul et al., [2021](#page-14-18)). However, the impact of the extracts on the accumulation of biogenic amines was different, depending on storage time and amine type (Özogul et al., [2021](#page-14-19)). Additionally, the carotenoid in C. vulgaris extracts, in particular, can be used to make bio-based plastic for food packaging. Lutein, αcarotene, and β‐carotene were isolated from C. vulgaris, and it was demonstrated that the addition of carotenoids to chitosan film considerably enhanced the film and demonstrated its suitability for use as a packaging material due to its antioxidative qualities (Şahin et al., [2019\)](#page-15-17). Hence, new investigations in terms of utilization of C. vulgaris extracts in bio‐based plastic are needed and how the extracts affect the mechanical and thermal properties of the bioplastics along with their effects on the shelf life of the packed products if they were used for packaging. Utilization the whole microalgae for food sector is commercially available and the most reasonable application of Chlorella sp. because the process is faster and easier than others which minimizes the overall cost (Enzing et al., [2014\)](#page-11-21). However, consumer demand significantly affects its market, and Spirulina dominates the market since 12,000 tons is produced yearly, while only 5000 tons is produced from Chlorella (Rani et al., [2018](#page-14-20)). Probably, understanding certain extracts from Chlorella that Spirulina does not or poorly have and correlating these extracts with health‐ boosting might raise Chlorella's market. The literature lacks information about the integration of a specific extract production with other productions since this process can reduce the overall cost leading to a better feasibility to produce an extract in industrial scale.

4.4 | Biocement

Biocement is a product innovation in the development of a bioprocess technology called Biocementing or Biocementation. Biocement refers to calcium carbonate (CaCO₃) deposits formed as a result of microbial activity in the systems that contain rich supplements in calcium ions. The major role of microorganisms in the precipitation of carbonate is due to their capability to produce an alkaline environment by many physiological actions (Ariyanti, [2012](#page-10-10)). The effects of different nitrogen sources and different

concentrations of sodium bicarbonate and carbon dioxide were investigated on C. vulgaris in terms of biomass concentration, biocement sedimentation rate, and productivity. According to the results, nitrate was favored over urea and ammonium, while the biocement efficiency was 90% and the biomass productivity was 490 mg L⁻¹ when calcium chloride (0.4 g L⁻¹), sodium bicarbonate $(2.5 g L⁻¹)$, and sodium nitrate $(1 g L⁻¹)$ were applied in the growth medium at the recommended proportions (Arabian, [2022](#page-10-17)). Further, the biomineralization capability of different species of microalgae in calcification media that contained sodium bicarbonate and calcium chloride dehydrate was investigated. Among the eight microalgae that have been examined, Synechocystis sp. had the highest calcium ion removal rate (0.70 mM day⁻¹), followed by C. vulgaris (0.40 mM day⁻¹) (Kavithraashree et al., [2022](#page-12-13)).

Calcium precipitation and C. vulgaris harvesting by flocculation showed a significant correlation. When the pH of the medium was adjusted to 11 for C. vulgaris harvesting, it was noticed that both calcium and magnesium were precipitated, and these precipitates can be utilized for further applications (Vandamme et al., [2012](#page-15-9)). Hence, the issue of pH dependency in microalgae harvesting (flocculation) can be employed or "invested" for biocement production or other applications related to calcium and/or magnesium. From what has been mentioned above, it can be considered that C. vulgaris as a promising candidate in civil engineering as well, and it would be more practical to treat wastewater that contains calcium ions and eventually utilize the biomass for different products, as mentioned previously. Prometheus Materials Inc. has unleashed the production of biocement from biomineralizing (the method through which living things' matrix absorbs mineral crystals) cyanobacteria and it was claimed that this cement is an alternative to Portland cement (Dreith, [2022\)](#page-11-22). Biocement mortars have a 90% lower thermal conductivity compared to conventional mortars, resulting in higher energy efficiency in buildings, reducing lifetime costs and $CO₂$ emissions associated with heating and cooling (Edwards, [2022\)](#page-11-23). Pyrolysis of the biomass was also suggested to improve the efficiency, and the addition of certain salts was suggested to be commercially beneficial (Maroušek et al., [2023](#page-13-14)). Yet, the optimization and biocement production by C. vulgaris needs to be investigated further in terms of calcium carbonate precipitation, the addition of the biomass effect into the produced biocement, and producing biocement by charring the biomass before and after oil extraction in two steps of biocement–biodiesel production. Finally, the price of the process must be investigated due to the lack of information

4.5 | Biodegradable bioplastics

4.5.1 | Polyhydroxyalkanoates

PHAs are biodegradable polyesters produced by bacteria (Kourmentza et al., [2017\)](#page-12-14), microalgae (Costa et al., [2019\)](#page-11-24), cyanobacteria (Afreen et al., [2021](#page-10-18)), and genetically engineered yeast (Gao et al., [2015](#page-12-15)). They serve as carbon and energy storage without toxic waste (Amstutz et al., [2019](#page-10-19)) and are fully recyclable. PHAs can be produced from various feedstocks, including wastes (Li & Wilkins, [2020\)](#page-13-15). However, microalgal biomass is considered the best due to its lower cost (Rahman et al., [2014](#page-14-21)). Chlorella fusca (Cassuriaga et al., [2018\)](#page-11-26), Chlorella pyrenoidosa (Das et al., 2018), Tetradesmus obliquus (Páblo Eugênio da Costa e & Laureen Michelle, [2022](#page-14-22)), Synechococcus subsalsus and Spirulina sp. (Costa et al., [2018](#page-11-27)) were reported for PHAs production, and it relays on the growth conditions, utilized media, $CO₂$ concentration, and nitrogen and phosphorus deficiency. For example, PHAs were produced by Synechococcus elongates; the production was 17% and 7% (w/w dry biomass) with nitrogen and phosphorus deficiency, respectively (Mendhulkar & Shetye, [2017\)](#page-13-16). Nostoc muscorum was also investigated under different growth conditions; PHAs production was 69% under phosphorus deficiency (Bhati & Mallick, [2015\)](#page-10-20) and 31% (w/w dry biomass) with acetate and propionate addition (Mallick et al., [2007](#page-13-17)). It was shown that C. vulgaris has the potential to produce PHAs by utilizing agro‐industry residue corn steep liquor (Páblo Eugênio da Costa e & Laureen Michelle, [2022](#page-14-22)). However, there is neither data about the exact concentration and productivity nor yield of PHAs. The most recent trend in PHA production by Chlorella sp. is the utilization of de‐oiled or de‐fatted biomass in the fermentation process to obtain the PHAs by microorganisms. Paracoccus sp. LL1 was employed to ferment glucose and Chlorella biomass to compare the produced PHAs along with carotenoids. Impressively, the obtained concentrations of PHAs and carotenoids were higher when the biomass was fermented; the concentrations that were obtained from biomass and sugar fermentation were $3.62 g L^{-1}$ PHAs and 11.7 mg L⁻¹ carotenoids and 1.48 g L⁻¹ PHAs and 6.08 mg L⁻¹ carotenoids, respectively (Khomlaem et al., [2020\)](#page-12-16). Moreover, the production of PHAs was improved when Chlorella biomass was fermented by Cupriavidus necator KCTC 2649, and Haloferax mediterranei DSM 1411. The highest PHAs production was found in C. necator KCTC 2649 with $7.51 \pm 0.20 \text{ g L}^{-1}$, 75% of dry cell weight, followed by 3.79 ± 0.03 g L⁻¹, 56% of dry cell weight in H. mediterranei DSM 1411. However, the carotenoids amount was decreased, and the maximum carotenoid content was 1.80 ± 0.16 mg L⁻¹ produced by H. mediterra-nei DSM 1411 (Khomlaem et al., [2021\)](#page-12-17). Despite the data available about de‐oiled biomass utilization to produce PHAs, the literature still lacks critical information about specifying the strain of Chlorella that has been grown to produce the biomass because the composition of the biomass is different. Moreover, specifying the strain of microalgae will be useful in biorefinery applications according to the desired final product such as biodiesel‐PHAs‐wastewater treatment simultaneously.

Polyhydroxybutyrate (PHB) is another polyester that belongs to the PHAs family and is produced by bacteria (McAdam et al., [2020](#page-13-18)) and microalgae (Robert & Iyer, [2018a](#page-14-23)). However, PHA has better chemical and physical properties and is comparable to petroleum‐ based plastics, whereas PHB has less elasticity, is very brittle, and its thermal properties are unsuitable for rigid products (Singh et al., [2015](#page-15-18)). PHB is a cost-effective, eco-friendly thermoplastic with similar properties to commercial polypropylene. It is less flexible than

traditional plastics and is completely biodegradable. PHB can be

0970290, 2024, 5, Downloaded from https:

wiley om/doi/10

.1002/bit.28666 by Izmir Ekonomi Univ

, Wiley

109/0204.3. Downloads the program with the control control control control of the control cont

Terms

and Conditions (https

//onlinelibrary.wiley

anditions

on Wiley Online Library

for rules of use; OP articles ing. governed by è

applicable Creati

Online Library on [17/04/2024]. See the

produced using algae, offering economic efficiency and low costs. Many microalgae species were investigated for PHB production; for instance, the maximum yield of PHB was 30% from 0.94 g L−¹ of C. sorokiniana biomass (Kumari et al., [2022](#page-13-19)), while it was up to 145.1 mg L^{-1} (17% of cell dry weight) under phosphate deficiency and limited $CO₂$ supply using N. muscorum (Haase et al., [2012\)](#page-12-18). Moreover, Spirulina sp. was described as a good candidate for PHB production; the yield was 21% of the cell dry weight in 15 days (Martins et al., [2017\)](#page-13-20). Recently, PHB was detected in C. vulgaris by its extraction from the biomass using hot chloroform (Robert & Iyer, [2018\)](#page-14-23) and it was quantified in a different study from 0.6 g of C. vulgaris biomass using mechanical (sonication) and chemical (NaClO) cell disruption methods, and the best yield was 37% (Setyorini & Dianursanti, [2021](#page-15-19)). Furthermore, C. vulgaris can be utilized in multiple production processes to make the process more sustainable and economical by utilizing the algal biomass completely without any production of algal waste. Recently, C. vulgaris was utilized to produce biodiesel and PHB by using a biorefinery; the microalgae were cultivated in wastewater, which makes the process more economical and sustainable by utilizing the algal biomass completely and without the production of any waste or residues. The authors utilized the biomass to extract the oil and then the de‐oiled biomass to produce PHB. The maximum lipid and PHB yield were 28 wt%, and 0.41 g g^{-1} , respectively (Arun et al., [2022\)](#page-10-21).

4.5.2 | Polylactic acid

PLA, an FDA‐approved thermoplastic monomer (Pines et al., [2008](#page-14-24)), is widely used in food handling (Marano et al., [2022\)](#page-13-21), 3D printing (Marșavina et al., [2022\)](#page-13-22) and promising for 4D printing (Lin et al., [2022\)](#page-13-23) and medical applications (Schätzlein et al., [2022](#page-15-20)). PLA is not toxic in solid form nor carcinogenic in the human body (Rogers, [2015\)](#page-14-25). When introduced into the human body, PLA hydrolyzes to alpha‐hydroxy acid then processed into the tricarboxylic acid cycle and excreted (Konta et al., [2017\)](#page-12-19). However, PLA can be toxic, if it is inhaled or absorbed into the skin or eyes as a vapor or liquid (Rogers, [2015](#page-14-25)). PLA can be derived from renewable sources like corn starch (Rogers, [2015](#page-14-25)), different microorganisms (Mehmood et al., [2023](#page-13-24)) including microalgae (Bussa et al., [2019\)](#page-11-28). The free‐lipid biomass from Nannochloropsis salina was utilized as a feedstock for Lactobacillus pentosus in fermentation process to produce lactic acid, and the results showed that lactic acid yield was 93% from 3 to 25 g L^{-1} sugar (Talukder et al., [2012\)](#page-15-21). In addition to that, lactic acid production by microalgal biomass fermentation was indicated to be practical on large scales when lactic acid was produced by Hydrodictyon reticulum fermentation which was carried out by Lactobacillus paracasei LA104. With a final concentration of 37.11 g L^{-1} and a productivity of 1.03 g L⁻¹ h⁻, the yield was 46 g/100 g H. Reticulum dry material (Nguyen et al., [2012](#page-14-26)). The potential of C. vulgaris to be utilized as a feedstock for fermentation was also studied. The results showed that C. vulgaris biomass contains a high source of carbohydrates, which can be utilized for lactic acid production (Agwa et al., [2022](#page-10-22)). Furthermore, it was noticed that C. vulgaris could change the enzymatic activity of Lactobacillus sp. When they were cocultured. The results showed higher production of L-lactic acid and lower Dlactic acid production, and this characteristic could be used to determine the final product (Ścieszka & Klewicka, [2020](#page-15-22)). However, more studies are required to understand the yield and productivity. The degradation rate of PLA depends on factors such as molecular weight, crystallinity, and stereochemistry (Tokiwa & Calabia, [2006](#page-15-23)). Thermal degradation, which occurs when 5% of the polymer is lost at 325°C and leaves no residue at 500°C (Sin et al., [2013](#page-15-24)), is not ecofriendly due to the emission of volatile compounds (Wojtyła et al., [2017](#page-16-12)). Enzymatic degradation, which takes several weeks to 24 months, is also possible by enzymes found in microorganisms such as protease (Seok et al., [2022](#page-15-25)), lipase (Satti et al., [2019](#page-15-26)), esterase (Mistry et al., [2022](#page-13-25)), and cutinase-like enzyme (Masaki et al., [2005](#page-13-26)), whereas other types of plastics take approximately 500 years (Kaushal et al., [2021](#page-12-20)). The rate of degradation is influenced by the polymer's structure and composition as well. D‐PLA degrading faster than L‐PLA (da Silva et al., [2018](#page-15-27)) and the addition of algal biomass to PLA accelerates the biodegradation of PLA‐based polymers (Kalita et al., [2021](#page-12-21)). However, there is no characterization of PLA/C. vulgaris in terms of biopolymer optimization and biodegradation analysis.

4.5.3 | PLA market size and share

PLA is relatively inexpensive when compared to other types of biodegradable bioplastics and has various mechanical advantages over others, making it a popular material. In 2019, the production

TABLE 2 A list of PLA products by different companies.

109/0204.3. Downloads the program with the control control control control of the control cont

and Conditions (https

wiley

Wiley Online Library

for rules of use; OP

ing.

governed by the

appicable Creativ

, Wiley Online Library on [17/04/2024]. See the Terms

10970290, 2024, 5, Downloaded from https:

elibrary.wiley.com/doi/10

.1002/bit.28666 by Izmir Ekonomi Univ

capacity of PLA was around 290,000 tons (Fortune Business Insights, [2021](#page-12-22)), and the global PLA market value was USD 1 billion in 2021 growing at CAGR of 12% from 2021 to 2026 (Markets and Markets, [2022](#page-13-27)), and expected to reach USD 2,306,708.2 thousand by 2028 (Fortune Business Insights, [2021\)](#page-12-22). In 2020, biodegradable plastics occupied 58% of the total plastic production globally, and among them, PLA had the largest share since its production capacity reached 19% and is expected to be 20% in 2025 from the total capacity of biodegradable plastic which is estimated to be 63% from plastic production (De Guzman, [2020](#page-12-23)). NatureWorks is the first company and the largest that produces PLA on a commercial scale in the world; it started in 2002, and in 2013, expanded its business and started to produce 150,000 metric tons annually. Furthermore, it has constructed a new facility in Thailand and is expected to be open in 2024; the facility will have an annual capacity of 75,000 tons of Ingeo® biopolymer (NatureWorks, [2021\)](#page-14-27). In line with NatureWorks company, Futerro also plans to set up an integrated factory in France to produce and recycle PLA with 75,000-ton capacity as an annual production (Futerro, [2022\)](#page-12-24). Some companies that produce PLA are mentioned in Table [2](#page-8-0) below. Packaging is the main application of PLA for consumer‐packaged goods, food service, and supermarket packaging. According to European Bioplastic, packaging accounted for 59% of biodegradable plastics produced in 2019. A fraction of this represents 0.5% of the total plastic packaging produced in biodegradable packaging. By the end of 2017, 164,000 biodegradable tons of packaging for food and food items were produced (Jia, [2020\)](#page-12-25). China is considered the largest PLA consumer, and it is expected to remain the world's largest PLA consumer over 2020‐2035, driven by new domestic capacity addition in the country, and also export demands (NexanTECA, [2021](#page-14-28)).

Abbreviations: n.d., referring to no data; PLA, polylactic acid.

FIGURE 3 Chlorella vulgaris applications. Bio-H₂, bio-hydrogen; CaCO₃, Calcium carbonate; PHA, polyhydroxyalkanoate; PLA, polylactic acid; W.W, wastewater.

4.5.4 | Plastic blends

Microalgae‐produced bioplastics have similar properties to petroleum‐based plastics, making them suitable for existing applications. They are biodegradable, making them eco‐friendly (Rahman & Miller, [2017\)](#page-14-6). Chlorella sp. Can be utilized to produce bioplastic because they have a high breaking strength due to their dense cell wall and high thermal stability (Cinar et al., [2020](#page-11-30)). Both compatibilizer and biomass treatment are effective for Chlorella‐based plastics. When the homogenizer was used for Chlorella powder treatment before mixing it with PVA, the results indicated that the treatment improved the tensile strength and elongation by 15.3 kgf/cm² and 100%, respectively (Sabathini et al., [2018\)](#page-15-29). Maleic anhydrates (MAs) compatibilizer showed tensile strength, elongation, elasticity, and homogeneity improvements, when it was incorporated with PVA‐ Chlorella plastic‐based (Dianursanti & Khalis, [2018\)](#page-11-31). Another composition was synthesized by blending Chlorella sp. with polyethylene (PE) through chemical modification of PE with MA. The result showed improved tensile strength by 40 wt % (Otsuki et al., [2004\)](#page-14-31). However, PE is not biodegradable. Therefore, biodegradable bioplastics like PLA are preferred. Although PLA has many benefits, as previously discussed, it also has significant disadvantages, including brittleness and low tear resistance (Kim et al., [2020](#page-12-28)). Therefore, there were many efforts including the incorporation of PLA with other plastics (Pivsa‐Art et al., [2016](#page-14-32)), plant biomass (Yoksan et al., [2022](#page-16-13)), or algal biomass (Liao et al., [2023](#page-13-30)) to improve the mechanical and thermal properties. Duckweed plant was incorporated with PLA/thermoplastic cassava starch (PLA/TPS) blend and the results showed improved Young's modulus, hardness, and tensile strength (Yoksan et al., [2022](#page-16-13)). PLA/algae blend was also investigated, and the results showed improved Young's modulus when macroalgae were incorporated with PLA. However, the strain and the tensile strength were decreased (Bulota & Budtova, [2015](#page-11-5)). Furthermore, cell disruption effect microalgae, namely spirulina sp. Blend with PLA was investigated as well, and it was demonstrated that the tensile strength was increased by 25% compared to raw spirulina (Liao et al., [2023\)](#page-13-30). Therefore, new investigations are required in terms of PLA/C. vulgaris blend along

with chemical composition analysis and biomass treatment impact the mechanical, thermal, and degradation properties of the biopolymer (bio-blend) since C. vulgaris was indicated for biorefinery application which leads to an economical process. Finally, a summary of overall C. vulgaris applications is mentioned in Figure [3](#page-9-0).

The process of producing bioplastics is expensive and, being a relatively new technology, less competitive than that of fossil‐based plastics (Shah & Gangadeen, [2023](#page-15-30)). That being said, given a stable price for fossil fuel, the price gap is anticipated to close considerably by 2030, considering a stable price of fossil fuel, and the price of fossil‐based plastics will be higher in a case of higher fossil fuel costs (Horvat et al., [2018\)](#page-12-29).

5 | CONCLUSION

C. vulgaris, easily cultivated and isolated from various sources, is a promising candidate for various bioprocesses and food supplements. Studying its unique extracts and their potential impact on human health could potentially boost its market, as consumer recognition directly influences its growth. Biodiesel production in industrial scale is the most feasible application, but several techno‐economic bottlenecks hinder its commercialization. These bottlenecks are linked to algae production and require processing as a side branch of a complex biorefinery process. Nontechnical factors like global fossil fuel costs, exchange rates, investor decisions, and governmental policies also affect biodiesel production and price, necessitating addressing these to improve green energy production. C. vulgaris has the potential for biohydrogen and biocement production; however, challenges related to hydrogen safety, operational cost, storage, and delivery hinder scaling. However, biocement and biomineralizing processes involving biomass and other biomaterials have not been extensively investigated. Microalgae‐based polymers are uncompetitive with fossil‐based. Probably, by valorization of de‐oiled biomass for biopolymer will be more practical if it is produced as a byproduct of other applications.

AUTHOR CONTRIBUTIONS

Mohammed Al‐Hammadi: Completed the literature review, analyzed the data, and wrote the manuscript. Mine Güngörmüşler: Designed and supervised the content and revised the manuscript. All authors read and approved the final manuscript.

ACKNOWLEDGMENTS

This research received no specific grant from any funding agency in the public, commercial, or not‐for‐profit sectors.

CONFLICT OF INTEREST STATEMENT

The authors declare no conflict of interest.

ORCID

Mohammed Al-Hammadi D <http://orcid.org/0009-0009-4317-3431> Mine Güngörmüşler **b** <http://orcid.org/0000-0002-0207-405X>

REFERENCES

- Abdo, S. M., Ahmed, E., El‐Enin, S. A., El Diwan, G., El‐Khatib, K. M., Ali, G. H., & Salah El Din, R. A. (2022). Algal fuel production by industry: Process simulation and economic assessment, Handbook of algal biofuels (pp. 635–652). Elsevier. [https://doi.org/10.1016/b978-](https://doi.org/10.1016/b978-0-12-823764-9.00029-7) [0-12-823764-9.00029-7](https://doi.org/10.1016/b978-0-12-823764-9.00029-7)
- Afreen, R., Tyagi, S., Singh, G. P., & Singh, M. (2021). Challenges and perspectives of polyhydroxyalkanoate production from microalgae/ cyanobacteria and bacteria as microbial factories: An assessment of hybrid biological system. Frontiers in Bioengineering and Biotechnology, 9, 1–14. <https://doi.org/10.3389/fbioe.2021.624885>
- Agrobiobase. (2013). Natureplast PLE 005. [http://www.agrobiobase.com/](http://www.agrobiobase.com/en/database/bioproducts/plastics-composites-rubber/natureplast-ple-005) [en/database/bioproducts/plastics-composites-rubber/natureplast](http://www.agrobiobase.com/en/database/bioproducts/plastics-composites-rubber/natureplast-ple-005)[ple-005](http://www.agrobiobase.com/en/database/bioproducts/plastics-composites-rubber/natureplast-ple-005)
- Aguda, R., Stelly, C., Fonseca, L., LeBoeuf, S., Massiha, S., Chistoserdov, A., Holmes, W. E., Hernandez, R., Zappi, M. E., & Revellame, E. D. (2023). Effect of macronutrient levels on Chlorella vulgaris cultivation for long duration spaceflights and space settlements. Acta Astronautica, 206, 206-217. [https://doi.org/10.1016/j.actaastro.](https://doi.org/10.1016/j.actaastro.2023.02.031) [2023.02.031](https://doi.org/10.1016/j.actaastro.2023.02.031)
- Agwa, O., Chukwunweike, C. A., & Ire, F. (2022). Production and optimization of lactic acid using Chlorella vulgaris as a source of fermentable sugar. Scientia Africana, 20(3), 119–140. [https://doi.](https://doi.org/10.4314/sa.v20i3.11) [org/10.4314/sa.v20i3.11](https://doi.org/10.4314/sa.v20i3.11)
- Akbari, M., Loganathan, N., Tavakolian, H., Mardani, A., & Streimikiene, D. (2021). The dynamic effect of micro‐structural shocks on private investment behavior. Acta Montanistica Slovaca, 26(1), 1–17. [https://](https://doi.org/10.46544/AMS.v26i1.01) doi.org/10.46544/AMS.v26i1.01
- Amaro, H. M., Salgado, E. M., Nunes, O. C., Pires, J. C. M., & Esteves, A. F. (2023). Microalgae systems—environmental agents for wastewater treatment and further potential biomass valorisation. Journal of Environmental Management, 337, 117678. [https://doi.org/10.1016/](https://doi.org/10.1016/j.jenvman.2023.117678) [j.jenvman.2023.117678](https://doi.org/10.1016/j.jenvman.2023.117678)
- Amstutz, V., Hanik, N., Pott, J., Utsunomia, C., & Zinn, M. (2019). Tailored biosynthesis of polyhydroxyalkanoates in chemostat cultures, Methods in enzymology (pp. 99–123). Academic Press. [https://doi.](https://doi.org/10.1016/bs.mie.2019.08.018) [org/10.1016/bs.mie.2019.08.018](https://doi.org/10.1016/bs.mie.2019.08.018)
- Anand, U., Dey, S., Parial, D., Federici, S., Ducoli, S., & Bolan, N. S., (2023). Algae and bacteria consortia for wastewater decontamination and transformation into biodiesel, bioethanol, biohydrogen, biofertilizers and animal feed: A review. Environmental Chemistry Letters, 21, 1585–1609. <https://doi.org/10.1007/s10311-023-01562-w>
- Antal, T. K., & Lindblad, P. (2005). Production of H2 by sulphur‐deprived cells of the unicellular Cyanobacteria Gloeocapsa alpicola and

Synechocystis sp. PCC 6803 during dark incubation with methane or at various extracellular pH. Journal of Applied Microbiology, 98(1), 114–120. <https://doi.org/10.1111/j.1365-2672.2004.02431.x>

- Arabian, D. (2022). Investigation of effective parameters on the productivity of biomass and bio‐cement as a soil improver from Chlorella vulgaris. Geomicrobiology Journal, 39(9), 781–790. [https://](https://doi.org/10.1080/01490451.2022.2078445) doi.org/10.1080/01490451.2022.2078445
- Ariana, R. (2016). Bioremediation of urban river wastewater using Chlorella vulgaris microalgae to generate biomass with potential for biodiesel production. Research, Society and Development, 2020, 1–23.
- Ariyanti, D. (2012). Feasibility of using microalgae for biocement production through biocementation. Journal of Bioprocessing & Biotechniques, 2(111). <https://doi.org/10.4172/2155-9821.1000111>
- Arun, J., Vigneshwar, S. S., Swetha, A., Gopinath, K. P., Basha, S., Brindhadevi, K., & Pugazhendhi, A. (2022). Bio‐based algal (Chlorella vulgaris) refinery on de‐oiled algae biomass cake: A study on biopolymer and biodiesel production. Science of the Total Environment, 816, 151579. [https://doi.org/10.1016/j.scitotenv.](https://doi.org/10.1016/j.scitotenv.2021.151579) [2021.151579](https://doi.org/10.1016/j.scitotenv.2021.151579)
- Asadi, P., Rad, H. A., & Qaderi, F. (2020). Lipid and biodiesel production by cultivation isolated strain Chlorella sorokiniana pa.91 and Chlorella vulgaris in dairy wastewater treatment plant effluents. Journal of Environmental Health Science and Engineering, 18(2), 573–585. <https://doi.org/10.1007/s40201-020-00483-y>
- Awaja, F., Daver, F., Kosior, E., & Cser, F. (2004). The effect of chain extension on the thermal behaviour and crystallinity of reactive extruded recycled pet. Journal of Thermal Analysis and Calorimetry, 78, 865–884. <https://doi.org/10.1007/s10973-005-0454-0>
- Ayatollahi, S. Z., Esmaeilzadeh, F., & Mowla, D. (2021). Integrated CO2 capture, nutrients removal and biodiesel production using Chlorella vulgaris. Journal of Environmental Chemical Engineering, 9(2), 104763. <https://doi.org/10.1016/j.jece.2020.104763>
- Barros, A. I., Gonçalves, A. L., Simões, M., & Pires, J. C. M. (2015). Harvesting techniques applied to microalgae: A review. Renewable and Sustainable Energy Reviews, 41, 1489–1500. [https://doi.org/10.](https://doi.org/10.1016/j.rser.2014.09.037) [1016/j.rser.2014.09.037](https://doi.org/10.1016/j.rser.2014.09.037)
- Batista, A. P., Ambrosano, L., Graça, S., Sousa, C., Marques, P. A. S. S., Ribeiro, B., Botrel, E. P., Castro Neto, P., & Gouveia, L. (2015). Combining urban wastewater treatment with biohydrogen production—An integrated microalgae‐based approach. Bioresource Technology, 184, 230–235. [https://doi.org/10.1016/j.biortech.2014.](https://doi.org/10.1016/j.biortech.2014.10.064) [10.064](https://doi.org/10.1016/j.biortech.2014.10.064)
- Beyerinick, M. W. (1890). Culturversuche mit Zoochlorellen, Lichenengonidien und anderen niederen Algen. Botanische Zeitung, 47, 725–785. [http://img.algaebase.org/pdf/AC100CF003338161AAoHt43C4207/](http://img.algaebase.org/pdf/AC100CF003338161AAoHt43C4207/35049.pdf) [35049.pdf](http://img.algaebase.org/pdf/AC100CF003338161AAoHt43C4207/35049.pdf)
- Bhati, R., & Mallick, N. (2015). Poly(3-hydroxybutyrate-co-3hydroxyvalerate) copolymer production by the diazotrophic cyanobacterium Nostoc muscorum Agardh: Process optimization and polymer characterization. Algal Research, 7, 78–85. [https://doi.org/](https://doi.org/10.1016/j.algal.2014.12.003) [10.1016/j.algal.2014.12.003](https://doi.org/10.1016/j.algal.2014.12.003)
- Bioplastics Magazine. (2020). FUTERRO launches the first fully integrated PLA plant in China. [https://www.bioplasticsmagazine.com/en/news/](https://www.bioplasticsmagazine.com/en/news/meldungen/20201022-FUTERRO-launches-the-first-fully-integrated-PLA-plant-in-China.php) [meldungen/20201022-FUTERRO-launches-the-first-fully](https://www.bioplasticsmagazine.com/en/news/meldungen/20201022-FUTERRO-launches-the-first-fully-integrated-PLA-plant-in-China.php)[integrated-PLA-plant-in-China.php](https://www.bioplasticsmagazine.com/en/news/meldungen/20201022-FUTERRO-launches-the-first-fully-integrated-PLA-plant-in-China.php)
- Bito, T., Okumura, E., Fujishima, M., & Watanabe, F. (2020). Potential of chlorella as a dietary supplement to promote human health. Nutrients, 12(9), 2524. <https://doi.org/10.3390/nu12092524>
- Bosma, R., Van Spronsen, W. A., Tramper, J., & Wijffels, R. H. (2003). Ultrasound, A new separation technique to harvest microalgae. Journal of Applied Phycology, 15(2–3), 143–153. [https://doi.org/10.](https://doi.org/10.1023/A:1023807011027) [1023/A:1023807011027](https://doi.org/10.1023/A:1023807011027)
- Branyikova, I., Prochazkova, G., Potocar, T., Jezkova, Z., & Branyik, T. (2018). Harvesting of microalgae by flocculation. Fermentation, 4(4), 93. <https://doi.org/10.3390/fermentation4040093>
- Brown, J. R. (2022). What is bioremediation, and how does it work (with examples)? Investopedia. [https://www.investopedia.com/terms/b/](https://www.investopedia.com/terms/b/bioremediation.asp#:~:text=Bioremediationisabranchof,,water,andotherenvironments) [bioremediation.asp#:~:text=Bioremediationisabranchof,%2Cwater%](https://www.investopedia.com/terms/b/bioremediation.asp#:~:text=Bioremediationisabranchof,,water,andotherenvironments) [2Candotherenvironments](https://www.investopedia.com/terms/b/bioremediation.asp#:~:text=Bioremediationisabranchof,,water,andotherenvironments)
- Bulota, M., & Budtova, T. (2015). PLA/algae composites: Morphology and mechanical properties. Composites, Part A: Applied Science and Manufacturing, 73, 109–115. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.compositesa.2015.03.001) [compositesa.2015.03.001](https://doi.org/10.1016/j.compositesa.2015.03.001)
- Bussa, M., Eisen, A., Zollfrank, C., & Röder, H. (2019). Life cycle assessment of microalgae products: State of the art and their potential for the production of polylactid acid. Journal of Cleaner Production, 213, 1299–1312. [https://doi.org/10.1016/j.jclepro.](https://doi.org/10.1016/j.jclepro.2018.12.048) [2018.12.048](https://doi.org/10.1016/j.jclepro.2018.12.048)
- Cassuriaga, A. P. A., Freitas, B. C. B., Morais, M. G., & Costa, J. A. V. (2018). Innovative polyhydroxybutyrate production by Chlorella fusca grown with pentoses. Bioresource Technology, 265, 456–463. [https://doi.](https://doi.org/10.1016/j.biortech.2018.06.026) [org/10.1016/j.biortech.2018.06.026](https://doi.org/10.1016/j.biortech.2018.06.026)
- CCAC. (2023). It's time to act on short‐lived climate pollutants for our food systems. Climate & Clean Air Coalition. [https://www.ccacoalition.](https://www.ccacoalition.org/news/its-time-act-short-lived-climate-pollutants-our-food-systems) [org/news/its-time-act-short-lived-climate-pollutants-our-food](https://www.ccacoalition.org/news/its-time-act-short-lived-climate-pollutants-our-food-systems)[systems](https://www.ccacoalition.org/news/its-time-act-short-lived-climate-pollutants-our-food-systems)
- Chaisson, C. (2021). Fossil fuel air pollution kills one in Five People. NRDC. [https://www.nrdc.org/stories/fossil-fuel-air-pollution-kills-one-five](https://www.nrdc.org/stories/fossil-fuel-air-pollution-kills-one-five-people#:~:text=Theestimated8.7millionpremature,thecombustionoffossilfuels)[people#:~:text=Theestima](https://www.nrdc.org/stories/fossil-fuel-air-pollution-kills-one-five-people#:~:text=Theestimated8.7millionpremature,thecombustionoffossilfuels) ted8.7millionpremature, [thecombustionoffossilfuels](https://www.nrdc.org/stories/fossil-fuel-air-pollution-kills-one-five-people#:~:text=Theestimated8.7millionpremature,thecombustionoffossilfuels)
- Chatham House. (2018). Making concrete change: Innovation in low‐carbon cement and concrete. [https://www.chathamhouse.org/2018/06/](https://www.chathamhouse.org/2018/06/making-concrete-change-innovation-low-carbon-cement-and-concrete) [making-concrete-change-innovation-low-carbon-cement-and](https://www.chathamhouse.org/2018/06/making-concrete-change-innovation-low-carbon-cement-and-concrete)[concrete](https://www.chathamhouse.org/2018/06/making-concrete-change-innovation-low-carbon-cement-and-concrete)
- Chen, J., Leng, L., Ye, C., Lu, Q., Addy, M., Wang, J., Liu, J., Chen, P., Ruan, R., & Zhou, W. (2018). A comparative study between fungal pellet- and spore-assisted microalgae harvesting methods for algae bioflocculation. Bioresource Technology, 259, 181-190. [https://doi.](https://doi.org/10.1016/j.biortech.2018.03.040) [org/10.1016/j.biortech.2018.03.040](https://doi.org/10.1016/j.biortech.2018.03.040)
- Chew, K. W., Yap, J. Y., Show, P. L., Suan, N. H., Juan, J. C., Ling, T. C., Lee, D. J., & Chang, J. S. (2017). Microalgae biorefinery: High value products perspectives. Bioresource Technology, 229, 53–62. [https://](https://doi.org/10.1016/j.biortech.2017.01.006) doi.org/10.1016/j.biortech.2017.01.006
- Chinh, N. T., & Hoang, T. (2022). Review polylactic acid: Synthesis, properties and technical and biomedical applications. Vietnam Journal of Science and Technology, 60(3), 283–313. [https://doi.org/](https://doi.org/10.15625/2525-2518/16721) [10.15625/2525-2518/16721](https://doi.org/10.15625/2525-2518/16721)
- CHO. (2020). More plastic is on the way: What it means for climate change. Columbia Climate School. [https://news.climate.columbia.edu/2020/](https://news.climate.columbia.edu/2020/02/20/plastic-production-climate-change/) [02/20/plastic-production-climate-change/](https://news.climate.columbia.edu/2020/02/20/plastic-production-climate-change/)
- Chovančíková, M., & Šimek, V. (2001). Effects of high‐fat and Chlorella vulgaris feeding on changes in lipid metabolism in mice. Biologia, 56(6), 661–666.
- Chozhavendhan, S., Karthigadevi, G., Praveenkumar, R., Aniskumar, M., Jayakumar, M., & Gurunathan, B. (2022). Potentials and challenges in biodiesel production from algae—Technological outlook, Biofuels and bioenergy: Opportunities and challenges (pp. 183–203). Elsevier. <https://doi.org/10.1016/B978-0-323-85269-2.00014-9>
- Cinar, S. O., Chong, Z. K., Kucuker, M. A., Wieczorek, N., Cengiz, U., & Kuchta, K. (2020). Bioplastic production from microalgae: A review. International Journal of Environmental Research and Public Health, 17(11), 1–21. <https://doi.org/10.3390/ijerph17113842>
- Costa, S. S., Miranda, A. L., Andrade, B. B., Assis, D. J., Souza, C. O., de Morais, M. G., Costa, J. A. V., & Druzian, J. I. (2018). Influence of nitrogen on growth, biomass composition, production, and properties of polyhydroxyalkanoates (PHAs) by microalgae. International Journal of Biological Macromolecules, 116, 552–562. [https://doi.org/](https://doi.org/10.1016/j.ijbiomac.2018.05.064) [10.1016/j.ijbiomac.2018.05.064](https://doi.org/10.1016/j.ijbiomac.2018.05.064)
- Costa, S. S., Miranda, A. L., de Morais, M. G., Costa, J. A. V., & Druzian, J. I. (2019). Microalgae as source of polyhydroxyalkanoates (PHAs)—A

review. International Journal of Biological Macromolecules, 536–547. <https://doi.org/10.1016/j.ijbiomac.2019.03.099>

- Das, S. K., Sathish, A., & Stanley, J. (2018). Production of biofuel and bioplastic from Chlorella Pyrenoidosa. Materials Today: Proceedings, 5(8), 16774–16781. <https://doi.org/10.1016/j.matpr.2018.06.020>
- Dewayanto, N., Adhi, K., Negara, N. A. K., Sadewo, B. R., Nisya, A. F., Prakoso, O., Sigit, U., Suyono, E. A., & Budiman, A. (2023). Study of low cost of microalgae chlorella sp. harvesting using cationic starch flocculation technique for biodiesel production. IOP Conference Series: Earth and Environmental Science, 1151(1), 012042. [https://doi.](https://doi.org/10.1088/1755-1315/1151/1/012042) [org/10.1088/1755-1315/1151/1/012042](https://doi.org/10.1088/1755-1315/1151/1/012042)
- Dianursanti, & Khalis, S. A. (2018). The effect of compatibilizer addition on Chlorella vulgaris microalgae utilization as a mixture for bioplastic. E3S Web of Conferences, 67, 2–6. [https://doi.org/10.1051/e3sconf/](https://doi.org/10.1051/e3sconf/20186703047) [20186703047](https://doi.org/10.1051/e3sconf/20186703047)
- Dreith, B. (2022). Prometheus materials uses algae‐based cement to make masonry blocks. Dezeen. [https://www.dezeen.com/2022/06/07/](https://www.dezeen.com/2022/06/07/prometheus-biocomposite-cement-blocks/) [prometheus-biocomposite-cement-blocks/](https://www.dezeen.com/2022/06/07/prometheus-biocomposite-cement-blocks/)
- Dvoretsky, D., Dvoretsky, S., Temnov, M., Akulinin, E., & Peshkova, E. (2016). Enhanced lipid extraction from microalgae Chlorella vulgaris biomass: Experiments, modelling, optimization. Chemical Engineering Transactions, 49, 175–180. <https://doi.org/10.3303/CET1649030>
- Dvoretsky, D. S., Akulinin, E., Temnov, M., Dvoretsky, A. D., EAkulinin, A., & Dvoretsky, S. (2016). Defining optimal conditions for Chlorella vulgaris microalgae biomass cell walls disruption in the process of biofuel production. Purification treatment of municipal wastewater using microalgae Chlorella vulgaris view project renewable energy sources and C. In 16th International Multidisciplinary Scientific GeoConference, SGEM 2016. <https://doi.org/10.5593/sgem2016B41>
- Edwards, P. (2022). Bio‐cement from Algae, global cement. [https://www.](https://www.globalcement.com/magazine/articles/1272-bio-cement-from-algae) [globalcement.com/magazine/articles/1272-bio-cement-from-algae](https://www.globalcement.com/magazine/articles/1272-bio-cement-from-algae)
- Ehman, N., & Area, M. C. (2021). Bioplastics are revolutionizing the packaging industry. BioResources, 16(3), 4663–4666. [https://doi.org/](https://doi.org/10.15376/biores.16.3.4663-4666) [10.15376/biores.16.3.4663-4666](https://doi.org/10.15376/biores.16.3.4663-4666)
- El‐fayoumy, E. A., Shanab, S. M. M., Gaballa, H. S., Tantawy, M. A., & Shalaby, E. A. (2021). Evaluation of antioxidant and anticancer activity of crude extract and different fractions of Chlorella vulgaris axenic culture grown under various concentrations of copper ions. BMC Complementary Medicine and Therapies, 21(1), 51. [https://doi.](https://doi.org/10.1186/s12906-020-03194-x) [org/10.1186/s12906-020-03194-x](https://doi.org/10.1186/s12906-020-03194-x)
- Enzing, C., Ploeg, M., Barbosa, M., & Sijtsma, L. (2014). Microalgae‐based products for the food and feed sector: An outlook for Europe. JRC Scientific and Policy Reports. <https://doi.org/10.2791/3339>
- Euglena. (2018). Euglena news. [https://www.euglena.jp/en/news/](https://www.euglena.jp/en/news/20181102en/) [20181102en/](https://www.euglena.jp/en/news/20181102en/)
- ExxonMobil. (2018). Advanced biofuels and algae research: Targeting the technical capability to produce 10000 barrels per day by 2025. [https://corporate.exxonmobil.com/climate-solutions/advanced](https://corporate.exxonmobil.com/climate-solutions/advanced-biofuels/advanced-biofuels-and-algae-research?print=true#Algaeforbiofuelsproduction)[biofuels/advanced-biofuels-and-algae-research?print=true#](https://corporate.exxonmobil.com/climate-solutions/advanced-biofuels/advanced-biofuels-and-algae-research?print=true#Algaeforbiofuelsproduction) [Algaeforbiofuelsproduction](https://corporate.exxonmobil.com/climate-solutions/advanced-biofuels/advanced-biofuels-and-algae-research?print=true#Algaeforbiofuelsproduction)
- Fasaei, F., Bitter, J. H., Slegers, P. M., & van Boxtel, A. J. B. (2018). Techno‐ economic evaluation of microalgae harvesting and dewatering systems. Algal Research, 31, 347–362. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.algal.2017.11.038) [algal.2017.11.038](https://doi.org/10.1016/j.algal.2017.11.038)
- Fayad, N., Yehya, T., Audonnet, F., & Vial, C. (2017). Harvesting of microalgae Chlorella vulgaris using electro‐coagulation‐flocculation in the batch mode. Algal Research, 25, 1–11. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.algal.2017.03.015) [algal.2017.03.015](https://doi.org/10.1016/j.algal.2017.03.015)
- Fazal, T., Rehman, M. S. U., Javed, F., Akhtar, M., Mushtaq, A., Hafeez, A., Alaud Din, A., Iqbal, J., Rashid, N., & Rehman, F. (2021). Integrating bioremediation of textile wastewater with biodiesel production using microalgae (Chlorella vulgaris). Chemosphere, 281, 130758. <https://doi.org/10.1016/j.chemosphere.2021.130758>
- Febrieni, V. N., Sedjati, S., & Yudiati, E. (2020). Optimization of light intensity on growth rate and total lipid content of Chlorella vulgaris.

IOP Conference Series: Earth and Environmental Science, 584(1), 012040. <https://doi.org/10.1088/1755-1315/584/1/012040>

- Feng, S., Ngo, H. H., Guo, W., Chang, S. W., Nguyen, D. D., Bui, X. T., Zhang, X., Ma, X. Y., & Hoang, B. N. (2023). Biohydrogen production, storage, and delivery: A comprehensive overview of current strategies and limitations. Chemical Engineering Journal, 471, 144669. <https://doi.org/10.1016/j.cej.2023.144669>
- Fortune Business Insights. (2021). Polylactic acid market size, share, & COVID−19 impact analysis, by application (packaging, textiles, comsumer goods, agriculture & horticulture, and others), and regional forecast, 2021‐2028. [https://www.fortunebusinessinsights.com/enquiry/](https://www.fortunebusinessinsights.com/enquiry/request-sample-book/polylactic-acid-pla-market-103429) [request-sample-book/polylactic-acid-pla-market-103429](https://www.fortunebusinessinsights.com/enquiry/request-sample-book/polylactic-acid-pla-market-103429)
- Freitas, H. R. (2017). Chlorella vulgaris as a source of essential fatty acids and micronutrients: A brief commentary. The Open Plant Science Journal, 10(1), 92-99. [https://doi.org/10.2174/](https://doi.org/10.2174/1874294701710010092) [1874294701710010092](https://doi.org/10.2174/1874294701710010092)
- Futerro. (2022). Futerro aims to set‐up a new fully integrated PLA biorefinery in Normandy, France. [https://www.futerro.com/news-media/futerro](https://www.futerro.com/news-media/futerro-aims-set-new-fully-integrated-pla-biorefinery-normandy-france#:~:text=Futerro,aBelgiancompanyand,productioncapacityof75,000tons)[aims-set-new-fully-integrated-pla-biorefinery-normandy-france#](https://www.futerro.com/news-media/futerro-aims-set-new-fully-integrated-pla-biorefinery-normandy-france#:~:text=Futerro,aBelgiancompanyand,productioncapacityof75,000tons) [:~:text=Futerro%2CaBelgiancompanyand,productioncapacityof75%](https://www.futerro.com/news-media/futerro-aims-set-new-fully-integrated-pla-biorefinery-normandy-france#:~:text=Futerro,aBelgiancompanyand,productioncapacityof75,000tons) [2C000tons](https://www.futerro.com/news-media/futerro-aims-set-new-fully-integrated-pla-biorefinery-normandy-france#:~:text=Futerro,aBelgiancompanyand,productioncapacityof75,000tons)
- Gao, C., Qi, Q., Madzak, C., & Lin, C. S. K. (2015). Exploring medium‐chain‐ length polyhydroxyalkanoates production in the engineered yeast Yarrowia lipolytica. Journal of Industrial Microbiology and Biotechnology, 42(9), 1255–1262. [https://doi.org/10.1007/s10295-](https://doi.org/10.1007/s10295-015-1649-y) [015-1649-y](https://doi.org/10.1007/s10295-015-1649-y)
- Gerulová, K., Kucmanová, A., Sanny, Z., Garaiová, Z., Seiler, E., Čaplovičová, M., Čaplovič, Ľ., & Palcut, M. (2022). Fe3O4‐PEI nanocomposites for magnetic harvesting of Chlorella vulgaris, Chlorella ellipsoidea, Microcystis aeruginosa, and Auxenochlorella protothecoides. Nanomaterials, 12(11), 1786. [https://doi.org/10.](https://doi.org/10.3390/nano12111786) [3390/nano12111786](https://doi.org/10.3390/nano12111786)
- Gotro, J. (2012). From corn to poly lactic acid (PLA): Fermentation in action, polymer innovation blog. [https://polymerinnovationblog.com/from](https://polymerinnovationblog.com/from-corn-to-poly-lactic-acid-pla-fermentation-in-action/)[corn-to-poly-lactic-acid-pla-fermentation-in-action/](https://polymerinnovationblog.com/from-corn-to-poly-lactic-acid-pla-fermentation-in-action/)
- De Guzman, D. (2015). Natureplast develops heat-resistant PLA resins. Green Chemicals Blog. [https://greenchemicalsblog.com/2015/03/](https://greenchemicalsblog.com/2015/03/10/natureplast-develops-heat-resistant-pla-resins/) [10/natureplast-develops-heat-resistant-pla-resins/](https://greenchemicalsblog.com/2015/03/10/natureplast-develops-heat-resistant-pla-resins/)
- De Guzman, D. (2020). Strong growth for bioplastics in 2020. Green Chemicals Blog. [https://greenchemicalsblog.com/2020/12/30/](https://greenchemicalsblog.com/2020/12/30/strong-growth-for-bioplastics-in-2020/) [strong-growth-for-bioplastics-in-2020/](https://greenchemicalsblog.com/2020/12/30/strong-growth-for-bioplastics-in-2020/)
- Haase, S. M., Huchzermeyer, B., & Rath, T. (2012). PHB accumulation in Nostoc muscorum under different carbon stress situations. Journal of Applied Phycology, 24(2), 157–162. [https://doi.org/10.1007/](https://doi.org/10.1007/s10811-011-9663-6) [s10811-011-9663-6](https://doi.org/10.1007/s10811-011-9663-6)
- Hidalgo, D., Martín‐Marroquín, J. M., & Corona, F. (2023). Metal‐based nanoadditives for increasing biomass and biohydrogen production in microalgal cultures: A review. Sustainable Chemistry and Pharmacy, 33, 101065. <https://doi.org/10.1016/j.scp.2023.101065>
- Hidayah Mat Yasin, N., Izzati Shafei, N., Hanani Rushan, N., Raihana Abu Sepian, N., & Mohd Said, F. (2019). The effect of microalgae harvesting on lipid for biodiesel production. Materials Today: Proceedings, 19, 1582–1590. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.matpr.2019.11.186) [matpr.2019.11.186](https://doi.org/10.1016/j.matpr.2019.11.186)
- Hong, J. W., Kim, O. H., Jo, S. W., Kim, H., Jeong, M. R., Park, K. M., Lee, K. I., & Yoon, H. S. (2016). Biochemical composition of a Korean domestic microalga Chlorella vulgaris KNUA027. Microbiology and Biotechnology Letters, 44(3), 400–407. <https://doi.org/10.4014/mbl.1512.12008>
- Horvat, D., Wydra, S., & Lerch, C. M. (2018). Modelling and simulating the dynamics of the European demand for bio‐based plastics. International Journal of Simulation Modelling, 17(3), 419–430. [https://doi.org/10.2507/IJSIMM17\(3\)435](https://doi.org/10.2507/IJSIMM17(3)435)
- Huang, Y., Wei, C., Liao, Q., Xia, A., Zhu, X., & Zhu, X. (2019). Biodegradable branched cationic starch with high C/N ratio for Chlorella vulgaris cells concentration: Regulating microalgae

flocculation performance by pH. Bioresource Technology, 276, 133–139. <https://doi.org/10.1016/J.BIORTECH.2018.12.072>

- IUCN. (2021). Marine plastic pollution. [https://www.iucn.org/resources/issues](https://www.iucn.org/resources/issues-brief/marine-plastic-pollution#:~:text=electronicsandagriculture.-,Over400milliontonsofplasticareproducedeveryyear,intheoceaneveryyear)[brief/marine-plastic-pollution#:~:text=electronicsandagriculture.-,](https://www.iucn.org/resources/issues-brief/marine-plastic-pollution#:~:text=electronicsandagriculture.-,Over400milliontonsofplasticareproducedeveryyear,intheoceaneveryyear) [Over400milliontonsofplasticareproducedeveryyear,intheoceaneveryyear](https://www.iucn.org/resources/issues-brief/marine-plastic-pollution#:~:text=electronicsandagriculture.-,Over400milliontonsofplasticareproducedeveryyear,intheoceaneveryyear)
- Jia, M. Z. (2020). Biodegradable plastics: Breaking down the facts. Production, composition and environmental impact (p. 54). [https://](https://www.greenpeace.org/static/planet4-eastasia-stateless/84075f56-biodegradable-plastics-report.pdf) [www.greenpeace.org/static/planet4-eastasia-stateless/84075f56](https://www.greenpeace.org/static/planet4-eastasia-stateless/84075f56-biodegradable-plastics-report.pdf) [biodegradable-plastics-report.pdf](https://www.greenpeace.org/static/planet4-eastasia-stateless/84075f56-biodegradable-plastics-report.pdf)
- Jiang, R., Qin, L., Feng, S., Huang, D., Wang, Z., & Zhu, S. (2021). The joint effect of ammonium and pH on the growth of Chlorella vulgaris and ammonium removal in artificial liquid digestate. Bioresource Technology, 325, 124690. [https://doi.org/10.1016/j.biortech.2021.](https://doi.org/10.1016/j.biortech.2021.124690) [124690](https://doi.org/10.1016/j.biortech.2021.124690)
- Love, J., & Bryant, J. A. (2017). Biofuels and Bioenergy (1st ed.). [https://doi.](https://doi.org/10.1002/9781118350553.ch6) [org/10.1002/9781118350553.ch6](https://doi.org/10.1002/9781118350553.ch6)
- Kalita, N. K., Damare, N. A., Hazarika, D., Bhagabati, P., Kalamdhad, A., & Katiyar, V. (2021). Biodegradation and characterization study of compostable PLA bioplastic containing algae biomass as potential degradation accelerator. Environmental Challenges, 3, 100067. <https://doi.org/10.1016/j.envc.2021.100067>
- Kaushal, J., Khatri, M., & Arya, S. K. (2021). Recent insight into enzymatic degradation of plastics prevalent in the environment: A mini‐review. Cleaner Engineering and Technology, 2, 100083. [https://doi.org/10.](https://doi.org/10.1016/j.clet.2021.100083) [1016/j.clet.2021.100083](https://doi.org/10.1016/j.clet.2021.100083)
- Kavithraashree, A., Rosfarizan, M., Siti Efliza, A., Joo Shun, T., & Mohd Shamzi, M. (2022). Bioprospecting microalgae with the capacity for inducing calcium carbonate biomineral precipitation. Asia‐Pacific Journal of Chemical Engineering, 17(3), e2767. [https://doi.](https://doi.org/10.1002/apj.2767) [org/10.1002/apj.2767](https://doi.org/10.1002/apj.2767)
- Khalaji, M., Hosseini, S. A., Ghorbani, R., Agh, N., Rezaei, H., Kornaros, M., & Koutra, E. (2021). Treatment of dairy wastewater by microalgae Chlorella vulgaris for biofuels production. Biomass Conversion and Biorefinery, 13(4), 3259–3265. [https://doi.org/10.1007/s13399-](https://doi.org/10.1007/s13399-021-01287-2) [021-01287-2](https://doi.org/10.1007/s13399-021-01287-2)
- Khetkorn, W., Rastogi, R. P., Incharoensakdi, A., Lindblad, P., Madamwar, D., Pandey, A., & Larroche, C. (2017). Microalgal hydrogen production—A review. Bioresource Technology, 243, 1194–1206. <https://doi.org/10.1016/j.biortech.2017.07.085>
- Kim, D. Y., Lee, J. B., Lee, D. Y., & Seo, K. H. (2020). Plasticization effect of poly(lactic acid) in the poly(butylene adipate–co–terephthalate) blown film for tear resistance improvement. Polymers, 12, 1904.
- Khomlaem, C., Aloui, H., Deshmukh, A. R., Yun, J. H., Kim, H. S., Napathorn, S. C., & Kim, B. S. (2020). Defatted Chlorella biomass as a renewable carbon source for polyhydroxyalkanoates and carotenoids co-production. Algal Research, 51, 102068. [https://doi.org/](https://doi.org/10.1016/j.algal.2020.102068) [10.1016/j.algal.2020.102068](https://doi.org/10.1016/j.algal.2020.102068)
- Khomlaem, C., Aloui, H., & Kim, B. S. (2021). Biosynthesis of polyhydroxyalkanoates from defatted chlorella biomass as an inexpensive substrate. Applied Sciences, 11(3), 1–11. [https://doi.org/10.3390/](https://doi.org/10.3390/app11031094) [app11031094](https://doi.org/10.3390/app11031094)
- Konta, A., García‐Piña, M., & Serrano, D. (2017). Personalised 3D printed medicines: Which techniques and polymers are more successful? Bioengineering, 4(4), 79. [https://doi.org/10.3390/bioengineering](https://doi.org/10.3390/bioengineering4040079) [4040079](https://doi.org/10.3390/bioengineering4040079)
- Kourmentza, C., Plácido, J., Venetsaneas, N., Burniol‐Figols, A., Varrone, C., Gavala, H. N., & Reis, M. A. M. (2017). Recent advances and challenges towards sustainable polyhydroxyalkanoate (PHA) production. Bioengineering, 4(2), 55. [https://doi.org/10.3390/](https://doi.org/10.3390/bioengineering4020055) [bioengineering4020055](https://doi.org/10.3390/bioengineering4020055)
- Kowalczyk, M., Piorkowska, E., Kulpinski, P., & Pracella, M. (2011). Mechanical and thermal properties of PLA composites with cellulose nanofibers and standard size fibers. Composites, Part A: Applied Science and Manufacturing, 42(10), 1509–1514. [https://doi.org/10.](https://doi.org/10.1016/j.compositesa.2011.07.003) [1016/j.compositesa.2011.07.003](https://doi.org/10.1016/j.compositesa.2011.07.003)
- Kumari, P., Ravi Kiran, B., & Venkata Mohan, S. (2022). Polyhydroxybutyrate production by Chlorella sorokiniana SVMIICT8 under nutrient‐ deprived mixotrophy. Bioresource Technology, 354, 127135. [https://](https://doi.org/10.1016/j.biortech.2022.127135) doi.org/10.1016/j.biortech.2022.127135
- Kurade, M. B., Kim, J. R., Govindwar, S. P., & Jeon, B. H. (2016). Insights into microalgae mediated biodegradation of diazinon by Chlorella vulgaris: Microalgal tolerance to xenobiotic pollutants and metabolism. Algal Research, 20, 126–134. [https://doi.org/10.1016/j.algal.](https://doi.org/10.1016/j.algal.2016.10.003) [2016.10.003](https://doi.org/10.1016/j.algal.2016.10.003)
- Lewin, R. A., & Andersen, R. A. (2022). algae. Encyclopedia Britannica. <https://www.britannica.com/science/algae>
- Li, M., & Wilkins, M. R. (2020). Recent advances in polyhydroxyalkanoate production: Feedstocks, strains and process developments. International Journal of Biological Macromolecules, 156, 691–703. <https://doi.org/10.1016/j.ijbiomac.2020.04.082>
- Li, S., Li, F., Zhu, X., Liao, Q., Chang, J. S., & Ho, S. H. (2022). Biohydrogen production from microalgae for environmental sustainability. Chemosphere, 291, 132717. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.chemosphere.2021.132717) [chemosphere.2021.132717](https://doi.org/10.1016/j.chemosphere.2021.132717)
- Liao, K., Grandgeorge, P., Jimenez, A. M., Nguyen, B. H., & Roumeli, E. (2023). Effects of mechanical cell disruption on the morphology and properties of spirulina‐PLA biocomposites. Sustainable Materials and Technologies, 36, e00591. [https://doi.org/10.1016/j.susmat.2023.](https://doi.org/10.1016/j.susmat.2023.e00591) [e00591](https://doi.org/10.1016/j.susmat.2023.e00591)
- Lin, C., Liu, L., Liu, Y., & Leng, J. (2022). 4D printing of shape memory polybutylene succinate/polylactic acid (PBS/PLA) and its potential applications. Composite Structures, 279, 114729. [https://doi.org/10.](https://doi.org/10.1016/J.COMPSTRUCT.2021.114729) [1016/J.COMPSTRUCT.2021.114729](https://doi.org/10.1016/J.COMPSTRUCT.2021.114729)
- Ma, W., Feng, C., Guan, F., Ma, D., & Cai, J. (2023). Effective Chlorella vulgaris biomass harvesting through sulfate and chloride flocculants. Journal of Marine Science and Engineering, 11(1), 47. [https://doi.org/](https://doi.org/10.3390/jmse11010047) [10.3390/jmse11010047](https://doi.org/10.3390/jmse11010047)
- Ma'mun, S., Wahyudi, A., & Raghdanesa, A. S. (2022). Growth rate measurements of Chlorella vulgaris in a photobioreactor by Neubauer‐improved counting chamber and densitometer. IOP Conference Series: Earth and Environmental Science, 963(1), 012015. <https://doi.org/10.1088/1755-1315/963/1/012015>
- Maazouz, A., & Lamnawar, K. (2011). Compounding and processing of biodegradable materials based on PLA for packaging applications: In Greening the 21st century materials world. Rheology and processing of recycled multilayer polymers view project coextrusion of multilayer polymers view project (April 2014). [https://www.researchgate.net/](https://www.researchgate.net/publication/261712108) [publication/261712108](https://www.researchgate.net/publication/261712108)
- Machineni, L., & Rao Anupoju, G. (2022). Review on valorization of lignocellulosic biomass for green plastics production: Sustainable and cleaner approaches. Sustainable Energy Technologies and Assessments, 53, 102698. <https://doi.org/10.1016/j.seta.2022.102698>
- Mallick, N., Gupta, S., Panda, B., & Sen, R. (2007). Process optimization for poly(3‐hydroxybutyrate‐co‐3‐hydroxyvalerate) co‐polymer production by Nostoc muscorum. Biochemical Engineering Journal, 37(2), 125–130. <https://doi.org/10.1016/j.bej.2007.04.002>
- Marano, S., Laudadio, E., Minnelli, C., & Stipa, P. (2022). Tailoring the barrier properties of PLA: A state-of-the-art review for food packaging applications. Polymers, 14(8), 1626. [https://doi.org/10.](https://doi.org/10.3390/polym14081626) [3390/polym14081626](https://doi.org/10.3390/polym14081626)
- Markets and Markets. (2022). Polylactic acid market by grade (thermoforming, extrusion, injection molding, blow molding), application (rigid thermoform, film & sheet, bottles), end‐use industry (packaging, consumer goods, agricultural, textile, biomedical) and region (2022‐ 2026). [https://www.marketsandmarkets.com/Market-Reports/](https://www.marketsandmarkets.com/Market-Reports/polylactic-acid-pla-market-29418964.html#:~:text=ThePolylacticAcidMarketis,drivingthemarketforPLA) polylactic-acid-pla-mar[ket-29418964.html#:~:text=](https://www.marketsandmarkets.com/Market-Reports/polylactic-acid-pla-market-29418964.html#:~:text=ThePolylacticAcidMarketis,drivingthemarketforPLA) [ThePolylacticAcidMarketis,drivingthemarketforPLA](https://www.marketsandmarkets.com/Market-Reports/polylactic-acid-pla-market-29418964.html#:~:text=ThePolylacticAcidMarketis,drivingthemarketforPLA)
- Maroušek, J. (2022). Review: Nanoparticles can change (bio)hydrogen competitiveness. Fuel, 328, 125318. [https://doi.org/10.1016/j.fuel.](https://doi.org/10.1016/j.fuel.2022.125318) [2022.125318](https://doi.org/10.1016/j.fuel.2022.125318)
- AL-HAMMADI and GÜNGÖRMÜŞLER \parallel 1499 BIOENGINEERIN
	- Maroušek, J., Gavurová, B., Strunecký, O., Maroušková, A., Sekar, M., & Marek, V. (2023). Techno‐economic identification of production factors threatening the competitiveness of algae biodiesel. Fuel, 344, 128056. <https://doi.org/10.1016/j.fuel.2023.128056>
	- Maroušek, J., Maroušková, A., Gavurová, B., & Minofar, B. (2023). Techno‐ economic considerations on cement substitute obtained from waste refining. Journal of Cleaner Production, 412, 137326. [https://doi.org/](https://doi.org/10.1016/j.jclepro.2023.137326) [10.1016/j.jclepro.2023.137326](https://doi.org/10.1016/j.jclepro.2023.137326)
	- Maroušek, J., Maroušková, A., Gavurová, B., Tuček, D., & Strunecký, O. (2023). Competitive algae biodiesel depends on advances in mass algae cultivation. Bioresource Technology, 374, 128802. [https://doi.](https://doi.org/10.1016/j.biortech.2023.128802) [org/10.1016/j.biortech.2023.128802](https://doi.org/10.1016/j.biortech.2023.128802)
	- Maroušek, J., Strunecký, O., Bartoš, V., & Vochozka, M. (2022). Revisiting competitiveness of hydrogen and algae biodiesel. Fuel, 328, 125317. <https://doi.org/10.1016/j.fuel.2022.125317>
	- Marșavina, L., Vălean, C., Mărghitaș, M., Linul, E., Razavi, N., Berto, F., & Brighenti, R. (2022). Effect of the manufacturing parameters on the tensile and fracture properties of FDM 3D‐printed PLA specimens. Engineering Fracture Mechanics, 274, 108766. [https://doi.org/10.](https://doi.org/10.1016/J.ENGFRACMECH.2022.108766) [1016/J.ENGFRACMECH.2022.108766](https://doi.org/10.1016/J.ENGFRACMECH.2022.108766)
	- Martins, R. G., Gonçalves, I. S., Morais, M. G., & Costa, J. A. V. (2017). New technologies from the bioworld: Selection of biopolymer‐producing microalgae. Polímeros, 27(4), 285–289. [https://doi.org/10.1590/](https://doi.org/10.1590/0104-1428.2375) [0104-1428.2375](https://doi.org/10.1590/0104-1428.2375)
	- Masaki, K., Kamini, N. R., Ikeda, H., & Iefuji, H. (2005). Cutinase‐like enzyme from the yeast Cryptococcus sp. strain S‐2 hydrolyzes polylactic acid and other biodegradable plastics. Applied and Environmental Microbiology, 71(11), 7548–7550. [https://doi.org/10.](https://doi.org/10.1128/AEM.71.11.7548-7550.2005) [1128/AEM.71.11.7548-7550.2005](https://doi.org/10.1128/AEM.71.11.7548-7550.2005)
	- MaterialDistrict. (2009). KARELINE. [https://materialdistrict.com/material/](https://materialdistrict.com/material/kareline/) [kareline/](https://materialdistrict.com/material/kareline/)
	- Mathimani, T., & Mallick, N. (2018). A comprehensive review on harvesting of microalgae for biodiesel—Key challenges and future directions. Renewable and Sustainable Energy Reviews, 91, 1103–1120. <https://doi.org/10.1016/j.rser.2018.04.083>
	- Ma, X., Zheng, H., Huang, H., Liu, Y., & Ruan, R. (2014). Effects of temperature and substrate concentration on lipid production by Chlorella vulgaris from enzymatic hydrolysates of lipid‐extracted microalgal biomass residues (LMBRs). Applied Biochemistry and Biotechnology, 174(4), 1631–1650. [https://doi.org/10.1007/](https://doi.org/10.1007/s12010-014-1134-5) [s12010-014-1134-5](https://doi.org/10.1007/s12010-014-1134-5)
	- McAdam, B., Brennan Fournet, M., McDonald, P., & Mojicevic, M. (2020). Production of polyhydroxybutyrate (PHB) and factors impacting its chemical and mechanical characteristics. Polymers, 12(12), 2908. <https://doi.org/10.3390/polym12122908>
	- Mehmood, A., Raina, N., Phakeenuya, V., Wonganu, B., & Cheenkachorn, K. (2023). The current status and market trend of polylactic acid as biopolymer: Awareness and needs for sustainable development. Materials Today: Proceedings, 72(6), 3049–3055. <https://doi.org/10.1016/J.MATPR.2022.08.387>
	- Mendhulkar, V. D., & Shetye, L. A. (2017). Synthesis of biodegradable polymer polyhydroxyalkanoate (PHA) in cyanobacteria Synechococcus elongates under mixotrophic nitrogen‐ and phosphate‐mediated stress conditions. Industrial Biotechnology, 13(2), 85–93. [https://doi.](https://doi.org/10.1089/ind.2016.0021) [org/10.1089/ind.2016.0021](https://doi.org/10.1089/ind.2016.0021)
	- Meticulous Research. (2022). Chlorella market. [https://www.](https://www.meticulousresearch.com/product/chlorella-market-5162) [meticulousresearch.com/product/chlorella-market-5162](https://www.meticulousresearch.com/product/chlorella-market-5162)
	- Metsoviti, M. N., Papapolymerou, G., Karapanagiotidis, I. T., & Katsoulas, N. (2020). Effect of light intensity and quality on growth rate and composition of Chlorella vulgaris. Plants, 9(1), 1–17. [https://](https://doi.org/10.3390/plants9010031) doi.org/10.3390/plants9010031
	- Mistry, A. N., Kachenchart, B., Wongthanaroj, A., Somwangthanaroj, A., & Luepromchai, E. (2022). Rapid biodegradation of high molecular weight semi‐crystalline polylactic acid at ambient temperature via enzymatic and alkaline hydrolysis by a defined bacterial consortium.

Polymer Degradation and Stability, 202, 110051. [https://doi.org/10.](https://doi.org/10.1016/j.polymdegradstab.2022.110051) [1016/j.polymdegradstab.2022.110051](https://doi.org/10.1016/j.polymdegradstab.2022.110051)

- de Morais, E. G., Sampaio, I. C. F., Gonzalez‐Flo, E., Ferrer, I., Uggetti, E., & García, J. (2023). Microalgae harvesting for wastewater treatment and resources recovery: A review. New Biotechnology, 78, 84–94. <https://doi.org/10.1016/j.nbt.2023.10.002>
- Mullarkey, M. (2017). Just 9% of discarded plastic recycled since 1950s. IEMA. [https://www.iema.net/articles/just-9-of-discarded-plastic](https://www.iema.net/articles/just-9-of-discarded-plastic-recycled-since-1950s#media-enquiry)[recycled-since-1950s#media-enquiry](https://www.iema.net/articles/just-9-of-discarded-plastic-recycled-since-1950s#media-enquiry)
- Nagy, V., Podmaniczki, A., Vidal‐Meireles, A., Tengölics, R., Kovács, L., Rákhely, G., Scoma, A., & Tóth, S. Z. (2018). Water‐splitting‐based, sustainable and efficient H2 production in green algae as achieved by substrate limitation of the Calvin‐Benson‐Bassham cycle. Biotechnology for Biofuels, 11(1), 69. <https://doi.org/10.1186/s13068-018-1069-0>
- NatureWorks. (2021). NatureWorks passes final authorization milestone for new fully integrated Ingeo PLA manufacturing plant in Thailand. [https://www.natureworksllc.com/News-and-Events/Press-](https://www.natureworksllc.com/News-and-Events/Press-Releases/2021/2021-08-09-NatureWorks-Final-Authorization-Ingeo-PLA-Plant-Thailand)[Releases/2021/2021-08-09-NatureWorks-Final-Authorization-](https://www.natureworksllc.com/News-and-Events/Press-Releases/2021/2021-08-09-NatureWorks-Final-Authorization-Ingeo-PLA-Plant-Thailand)[Ingeo-PLA-Plant-Thailand](https://www.natureworksllc.com/News-and-Events/Press-Releases/2021/2021-08-09-NatureWorks-Final-Authorization-Ingeo-PLA-Plant-Thailand)
- NatureWorks. (2023). We make Ingeo, a new material for plastics & fibers with unique properties that all begin with greenhouse gases. [https://](https://www.natureworksllc.com/) www.natureworksllc.com/
- NexanTECA. (2021). Global Polylactic Acid (PLA) market snapshot. [https://](https://www.nexanteca.com/blog/202106/global-polylactic-acid-pla-market-snapshot#:~:text=ThelargestmarketforPLA,domesticandalsoexportdemand) [www.nexanteca.com/blog/202106/global-polylactic-acid-pla](https://www.nexanteca.com/blog/202106/global-polylactic-acid-pla-market-snapshot#:~:text=ThelargestmarketforPLA,domesticandalsoexportdemand)[market-snapshot#:~:text=ThelargestmarketforPLA,](https://www.nexanteca.com/blog/202106/global-polylactic-acid-pla-market-snapshot#:~:text=ThelargestmarketforPLA,domesticandalsoexportdemand) [domesticandalsoexportdemand](https://www.nexanteca.com/blog/202106/global-polylactic-acid-pla-market-snapshot#:~:text=ThelargestmarketforPLA,domesticandalsoexportdemand)
- Nguyen, C. M., Kim, J. S., Hwang, H. J., Park, M. S., Choi, G. J., Choi, Y. H., Jang, K. S., & Kim, J. C. (2012). Production of l‐lactic acid from a green microalga, Hydrodictyon reticulum, by Lactobacillus paracasei LA104 isolated from the traditional Korean food, makgeolli. Bioresource Technology, 110, 552–559. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.biortech.2012.01.079) [biortech.2012.01.079](https://doi.org/10.1016/j.biortech.2012.01.079)
- Otsuki, T., Zhang, F., Kabeya, H., & Hirotsu, T. (2004). Synthesis and tensile properties of a novel composite of chlorella and polyethylene. Journal of Applied Polymer Science, 92(2), 812–816. [https://](https://doi.org/10.1002/app.13650) doi.org/10.1002/app.13650
- Özogul, İ., Kuley, E., Durmus, M., Özogul, Y., & Polat, A. (2021). The effects of microalgae (Spirulina platensis and Chlorella vulgaris) extracts on the quality of vacuum packaged sardine during chilled storage. Journal of Food Measurement and Characterization, 15(2), 1327–1340. [https://doi.](https://doi.org/10.1007/s11694-020-00729-1) [org/10.1007/s11694-020-00729-1](https://doi.org/10.1007/s11694-020-00729-1)
- Özogul, İ., Kuley, E., Ucar, Y., Yazgan, H., & Özogul, Y. (2021). Inhibitory impacts of Spirulina platensis and Chlorella vulgaris extracts on biogenic amine accumulation in sardine fillets. Food Bioscience, 41, 101087. <https://doi.org/10.1016/j.fbio.2021.101087>
- Páblo Eugênio da Costa e, S., & Laureen Michelle, H. (2022). Obtainment of polyhydroxyalkanoates (PHAs) from microalgae supplemented with Agro-Industry residue corn steep liquor. Journal of Botany Research, 5(1), 138–140. <https://doi.org/10.36959/771/571>
- Panahi, Y., Darvishi, B., Jowzi, N., Beiraghdar, F., & Sahebkar, A. (2016). Chlorella vulgaris: A multifunctional dietary supplement with diverse medicinal properties. Current Pharmaceutical Design, 22(2), 164–173. <https://doi.org/10.2174/1381612822666151112145226>
- Panahi, Y., Pishgoo, B., Jalalian, H. R., Mohammadi, E., Taghipour, H. R., Sahebkar, A., & Abolhasani, E. (2012). Investigation of the effects of Chlorella vulgaris as an adjunctive therapy for dyslipidemia: Results of a randomised open‐label clinical trial. Nutrition & Dietetics, 69(1), 13–19. <https://doi.org/10.1111/j.1747-0080.2011.01569.x>
- Pavolová, H., Bakalár, T., Kyšeľa, K., Klimek, M., Hajduová, Z., & Zawada, M. (2021). The analysis of investment into industries based on portfolio managers. Acta Montanistica Slovaca, 26(1), 161–170. <https://doi.org/10.46544/AMS.v26i1.14>
- Pines, E., Ph, D., Device, S., & Dressing, N. (2008). 6. Substantial equivalence summary. [https://www.accessdata.fda.gov/cdrh_docs/](https://www.accessdata.fda.gov/cdrh_docs/pdf8/K082276.pdf) [pdf8/K082276.pdf](https://www.accessdata.fda.gov/cdrh_docs/pdf8/K082276.pdf)
- Pivsa‐Art, S., Kord‐Sa‐Ard, J., Pivsa‐Art, W., Wongpajan, R., O‐Charoen, N., Pavasupree, S., & Hamada, H. (2016). Effect of compatibilizer on PLA/PP blend for injection molding. Energy Procedia, 89, 353–360. <https://doi.org/10.1016/j.egypro.2016.05.046>
- Prochazkova, G., Podolova, N., Safarik, I., Zachleder, V., & Branyik, T. (2013). Physicochemical approach to freshwater microalgae harvesting with magnetic particles. Colloids and Surfaces B: Biointerfaces, 112, 213–218. <https://doi.org/10.1016/j.colsurfb.2013.07.053>
- Pugazhendhi, A., Shobana, S., Bakonyi, P., Nemestóthy, N., Xia, A., Banu J, R., & Kumar, G. (2019). A review on chemical mechanism of microalgae flocculation via polymers. Biotechnology Reports, 21, e00302. <https://doi.org/10.1016/j.btre.2018.e00302>
- Rahman, A., Anthony, R. J., Sathish, A., Sims, R. C., & Miller, C. D. (2014). Effects of wastewater microalgae harvesting methods on polyhydroxybutyrate production. Bioresource Technology, 156, 364–367. <https://doi.org/10.1016/J.BIORTECH.2014.01.034>
- Rahman, A., & Miller, C. D. (2017). Microalgae as a source of bioplastics, Algal green chemistry: Recent progress in biotechnology. Elsevier B.V. <https://doi.org/10.1016/B978-0-444-63784-0.00006-0>
- Rahman, M. H., & Bhoi, P. R. (2021). An overview of non‐biodegradable bioplastics. Journal of Cleaner Production, 294, 126218. [https://doi.](https://doi.org/10.1016/j.jclepro.2021.126218) [org/10.1016/j.jclepro.2021.126218](https://doi.org/10.1016/j.jclepro.2021.126218)
- Rajagopalan, S., & Landrigan, P. J. (2021). Pollution and the heart. New England Journal of Medicine, 385(20), 1881–1892. [https://doi.org/](https://doi.org/10.1056/nejmra2030281) [10.1056/nejmra2030281](https://doi.org/10.1056/nejmra2030281)
- Rani, K., Sandal, N., & Sahoo, P. K. (2018). A comprehensive review on chlorella‐its composition, health benefits, market and regulatory scenario. The Pharma Innovation Journal, 7(7), 584–589. [https://](https://www.thepharmajournal.com) www.thepharmajournal.com
- Rashid, N., Lee, K., & Mahmood, Q. (2011). Bio‐hydrogen production by Chlorella vulgaris under diverse photoperiods. Bioresource Technology, 102(2), 2101–2104. [https://doi.org/10.1016/j.biortech.2010.](https://doi.org/10.1016/j.biortech.2010.08.032) [08.032](https://doi.org/10.1016/j.biortech.2010.08.032)
- Rashid, N., Rehman, S. U., & Han, J. I. (2013). Rapid harvesting of freshwater microalgae using chitosan. Process Biochemistry, 48(7), 1107–1110. <https://doi.org/10.1016/j.procbio.2013.04.018>
- Rendón, S. M., Roldan, G. J., & Voroney, R. P. (2013). Effect of carbon dioxide concentration on the growth response of Chlorella vulgaris under four different LED illumination. International Journal of Biotechnology for Wellness Industries, 2(3), 125. [https://doi.org/10.](https://doi.org/10.6000/1927-3037.2013.02.03.3) [6000/1927-3037.2013.02.03.3](https://doi.org/10.6000/1927-3037.2013.02.03.3)
- Renewable Carbon News. (2012). RTP Company expands bioplastic products to include impact‐modified PLA compounds. [https://](https://renewable-carbon.eu/news/rtp-company-expands-bioplastic-products-to-include-impact-modified-pla-compounds/) [renewable-carbon.eu/news/rtp-company-expands-bioplastic](https://renewable-carbon.eu/news/rtp-company-expands-bioplastic-products-to-include-impact-modified-pla-compounds/)[products-to-include-impact-modified-pla-compounds/](https://renewable-carbon.eu/news/rtp-company-expands-bioplastic-products-to-include-impact-modified-pla-compounds/)
- Robert, R., & Iyer, P. R. (2018). Isolation and optimization of PHB (Poly‐β‐ hydroxybutyrate) based biodegradable plastics from Chlorella vulgaris. Journal of Bioremediation & Biodegradation, 9(2), 2–5. [https://doi.](https://doi.org/10.4172/2155-6199.1000433) [org/10.4172/2155-6199.1000433](https://doi.org/10.4172/2155-6199.1000433)
- Roberts, D. (2020). Big Oil's hopes are pinned on plastics. It won't end well. Vox. [https://www.vox.com/energy-and-environment/21419505/](https://www.vox.com/energy-and-environment/21419505/oil-gas-price-plastics-peak-climate-change) [oil-gas-price-plastics-peak-climate-change](https://www.vox.com/energy-and-environment/21419505/oil-gas-price-plastics-peak-climate-change)
- Rochaix, J.‐D. (2016). The dynamics of the photosynthetic apparatus in algae. Applied Photosynthesis—New Progress. [https://doi.org/10.](https://doi.org/10.5772/62261) [5772/62261](https://doi.org/10.5772/62261)
- Rogers, T. (2015). Everything you need to know about polylactic acid (PLA). Creative Mechanisms. [https://www.creativemechanisms.com/blog/](https://www.creativemechanisms.com/blog/learn-about-polylactic-acid-pla-prototypes) [learn-about-polylactic-acid-pla-prototypes](https://www.creativemechanisms.com/blog/learn-about-polylactic-acid-pla-prototypes)
- Rubenstein, M. (2012). Emissions from the cement industry. Columbia Climate School. [https://news.climate.columbia.edu/2012/05/09/](https://news.climate.columbia.edu/2012/05/09/emissions-from-the-cement-industry/#:~:text=Cementmanufacturing%20is%20highly%20energy,atonofCO2) [emissions-from-the-cem](https://news.climate.columbia.edu/2012/05/09/emissions-from-the-cement-industry/#:~:text=Cementmanufacturing%20is%20highly%20energy,atonofCO2) ent-industry/#:~:text= [Cementmanufacturing%20is%20highly%20energy,atonofCO2](https://news.climate.columbia.edu/2012/05/09/emissions-from-the-cement-industry/#:~:text=Cementmanufacturing%20is%20highly%20energy,atonofCO2)
- Ruiz‐Marin, A., Canedo‐López, Y., & Chávez‐Fuentes, P. (2020). Biohydrogen production by Chlorella vulgaris and Scenedesmus obliquus immobilized cultivated in artificial wastewater under different light

e Commons

quality. AMB Express, 10(1), 191. [https://doi.org/10.1186/s13568-](https://doi.org/10.1186/s13568-020-01129-w) [020-01129-w](https://doi.org/10.1186/s13568-020-01129-w)

- Sabathini, H. A., Windiani, L., Dianursanti, & Gozan, M. (2018). Mechanical physical properties of Chlorella‐PVA based bioplastic with ultrasonic homogenizer. E3S Web of Conferences, 67, 3-7. [https://doi.org/10.](https://doi.org/10.1051/e3sconf/20186703046) [1051/e3sconf/20186703046](https://doi.org/10.1051/e3sconf/20186703046)
- Safi, C., Zebib, B., Merah, O., Pontalier, P. Y., & Vaca‐Garcia, C. (2014). Morphology, composition, production, processing and applications of Chlorella vulgaris: A review. Renewable and Sustainable Energy Reviews, 35, 265–278. <https://doi.org/10.1016/j.rser.2014.04.007>
- Şahin, S., Tsiqah Binti Mohd Nasir, N., Erken, I., Elibol Çakmak, Z., & Çakmak, T. (2019). Antioxidant composite films with chitosan and carotenoid extract from Chlorella vulgaris: Optimization of ultrasonic‐ assisted extraction of carotenoids and surface characterization of chitosan films. Materials Research Express, 6(9), 095404. [https://doi.](https://doi.org/10.1088/2053-1591/ab2def) [org/10.1088/2053-1591/ab2def](https://doi.org/10.1088/2053-1591/ab2def)
- Sahu, O. (2014). Reduction of organic and inorganic pollutant from waste water by algae. International Letters of Natural Sciences, 13(1), 1–8. <https://doi.org/10.56431/p-8aq47u>
- Salvia, S., Mirzah, M., Marlida, Y., & Purwati, E. (2014). The optimizing of growth and quality of Chlorella vulgaris as ASUH feed supplement for broiler. International Journal on Advanced Science, Engineering and Information Technology, 4(4), 294. [https://doi.org/10.18517/ijaseit.4.](https://doi.org/10.18517/ijaseit.4.4.421) [4.421](https://doi.org/10.18517/ijaseit.4.4.421)
- Satti, S. M., Abbasi, A. M., Salahuddin, Rana, Q. A., Marsh, T. L., Auras, R., Hasan, F., Badshah, M., Farman, M., & Shah, A. A. (2019). Statistical optimization of lipase production from Sphingobacterium sp. strain S2 and evaluation of enzymatic depolymerization of Poly(lactic acid) at mesophilic temperature. Polymer Degradation and Stability, 160, 1–13. <https://doi.org/10.1016/j.polymdegradstab.2018.11.030>
- Save on Energy. (2022). American food production requires more energy than you'd think. [https://www.saveonenergy.com/resources/food](https://www.saveonenergy.com/resources/food-production-requires-energy/)[production-requires-energy/](https://www.saveonenergy.com/resources/food-production-requires-energy/)
- Savvidou, M. G., Dardavila, M. M., Georgiopoulou, I., Louli, V., Stamatis, H., Kekos, D., & Voutsas, E. (2021). Optimization of microalga Chlorella vulgaris magnetic harvesting. Nanomaterials, 11(6), 1614. [https://doi.](https://doi.org/10.3390/nano11061614) [org/10.3390/nano11061614](https://doi.org/10.3390/nano11061614)
- Schätzlein, E., Kicker, C., Söhling, N., Ritz, U., Neijhoft, J., Henrich, D., Frank, J., Marzi, I., & Blaeser, A. (2022). 3D‐Printed PLA‐bioglass scaffolds with controllable calcium release and MSC adhesion for bone tissue engineering. Polymers, 14(12), 2389. [https://doi.org/10.](https://doi.org/10.3390/polym14122389) [3390/polym14122389](https://doi.org/10.3390/polym14122389)
- Ścieszka, S., & Klewicka, E. (2020). Influence of the microalga Chlorella vulgaris on the growth and metabolic activity of Lactobacillus spp. bacteria. Foods, 9(7), 959. <https://doi.org/10.3390/foods9070959>
- Scragg, A. H., Morrison, J., & Shales, S. W. (2003). The use of a fuel containing Chlorella vulgaris in a diesel engine. Enzyme and Microbial Technology, 33(7), 884–889. [https://doi.org/10.1016/j.enzmictec.](https://doi.org/10.1016/j.enzmictec.2003.01.001) [2003.01.001](https://doi.org/10.1016/j.enzmictec.2003.01.001)
- Sekar, M., Praveen Kumar, T. R., Selva Ganesh Kumar, M., Vaníčková, R., & Maroušek, J. (2021). Techno‐economic review on short‐term anthropogenic emissions of air pollutants and particulate matter. Fuel, 305, 121544. <https://doi.org/10.1016/j.fuel.2021.121544>
- Seok, J. H., Enomoto, Y., & Iwata, T. (2022). Synthesis of paramylon ester‐ graft‐PLA copolymers and its two‐step enzymatic degradation by proteinase K and β‐1,3‐glucanase. Polymer Degradation and Stability, 197, 109855. [https://doi.org/10.1016/J.POLYMDEGRADSTAB.](https://doi.org/10.1016/J.POLYMDEGRADSTAB.2022.109855) [2022.109855](https://doi.org/10.1016/J.POLYMDEGRADSTAB.2022.109855)
- Setyorini, A. I., & Dianursanti, D. (2021). Isolation and optimization of polyhydroxybutyrate from Chlorella vulgaris using NaClO and ultrasonication aided chemical pretreatment methods. AIP Conference Proceedings, 2344(1). [https://doi.org/10.1063/5.](https://doi.org/10.1063/5.0047562) [0047562](https://doi.org/10.1063/5.0047562)
- Shagun. (2023). Food production accounts for 15% fossil fuel use, as big oil locks in dependence on petroleum‐based pesticides. Down To Earth.

[https://www.downtoearth.org.in/news/agriculture/food](https://www.downtoearth.org.in/news/agriculture/food-production-accounts-for-15-fossil-fuel-use-as-big-oil-locks-in-dependence-on-petroleum-based-pesticides-92618)[production-accounts-for-15-fossil-fuel-use-as-big-oil-locks-in](https://www.downtoearth.org.in/news/agriculture/food-production-accounts-for-15-fossil-fuel-use-as-big-oil-locks-in-dependence-on-petroleum-based-pesticides-92618)[dependence-on-petroleum-based-pesticides-92618](https://www.downtoearth.org.in/news/agriculture/food-production-accounts-for-15-fossil-fuel-use-as-big-oil-locks-in-dependence-on-petroleum-based-pesticides-92618)

- Shah, K. U., & Gangadeen, I. (2023). Integrating bioplastics into the US plastics supply chain: Towards a policy research agenda for the bioplastic transition. Frontiers in Environmental Science, 11, 245846. <https://doi.org/10.3389/fenvs.2023.1245846>
- Sherafati, N., Bideshki, M. V., Behzadi, M., Mobarak, S., Asadi, M., & Sadeghi, O. (2022). Effect of supplementation with Chlorella vulgaris on lipid profile in adults: A systematic review and dose‐response meta‐analysis of randomized controlled trials. Complementary Therapies in Medicine, 66, 102822. [https://doi.org/10.1016/J.CTIM.](https://doi.org/10.1016/J.CTIM.2022.102822) [2022.102822](https://doi.org/10.1016/J.CTIM.2022.102822)
- da Silva, D., Kaduri, M., Poley, M., Adir, O., Krinsky, N., Shainsky‐Roitman, J., & Schroeder, A. (2018). Biocompatibility, biodegradation and excretion of polylactic acid (PLA) in medical implants and theranostic systems. Chemical Engineering Journal, 340, 9–14. [https://doi.org/](https://doi.org/10.1016/j.cej.2018.01.010) [10.1016/j.cej.2018.01.010](https://doi.org/10.1016/j.cej.2018.01.010)
- Sin, L. T., Rahmat, A. R., & Rahman, W. A. W. A. (2013). Degradation and stability of poly(lactic Acid), Polylactic acid (pp. 247–299). [https://](https://doi.org/10.1016/b978-1-4377-4459-0.00007-x) doi.org/10.1016/b978-1-4377-4459-0.00007-x
- Singh, M., Kumar, P., Ray, S., & Kalia, V. C. (2015). Challenges and opportunities for customizing polyhydroxyalkanoates. Indian Journal of Microbiology, 55(3), 235–249. [https://doi.org/10.1007/s12088-](https://doi.org/10.1007/s12088-015-0528-6) [015-0528-6](https://doi.org/10.1007/s12088-015-0528-6)
- Skyquest. (2022). Global Chlorella Market. [https://www.skyquestt.com/](https://www.skyquestt.com/report/chlorella-market#:~:text=GlobalChlorella%20Market%20Insights,period(2023%E2%80%902030)) [report/chlorella-market#:~:text=GlobalChlorella%20Market%](https://www.skyquestt.com/report/chlorella-market#:~:text=GlobalChlorella%20Market%20Insights,period(2023%E2%80%902030)) [20Insights,period\(2023-2030\)](https://www.skyquestt.com/report/chlorella-market#:~:text=GlobalChlorella%20Market%20Insights,period(2023%E2%80%902030))
- Stávková, J., & Maroušek, J. (2021). Novel sorbent shows promising financial results on P recovery from sludge water. Chemosphere, 276, 130097. <https://doi.org/10.1016/j.chemosphere.2021.130097>
- Sudesh, K., & Iwata, T. (2008). Sustainability of biobased and biodegradable plastics. CLEAN – Soil, Air, Water, 36(5–6), 433–442. <https://doi.org/10.1002/clen.200700183>
- Talukder, M. M. R., Das, P., & Wu, J. C. (2012). Microalgae (Nannochloropsis salina) biomass to lactic acid and lipid. Biochemical Engineering Journal, 68, 109–113. <https://doi.org/10.1016/j.bej.2012.07.001>
- Tokiwa, Y., & Calabia, B. P. (2006). Biodegradability and biodegradation of poly(lactide. Applied Microbiology and Biotechnology, 72(2), 244–251. <https://doi.org/10.1007/s00253-006-0488-1>
- Touloupakis, E., Faraloni, C., Silva Benavides, A. M., Masojídek, J., & Torzillo, G. (2021). Sustained photobiological hydrogen production by Chlorella vulgaris without nutrient starvation. International Journal of Hydrogen Energy, 46(5), 3684–3694. [https://doi.org/10.1016/J.](https://doi.org/10.1016/J.IJHYDENE.2020.10.257) [IJHYDENE.2020.10.257](https://doi.org/10.1016/J.IJHYDENE.2020.10.257)
- Ummalyma, S. B., Mathew, A. K., Pandey, A., & Sukumaran, R. K. (2016). Harvesting of microalgal biomass: Efficient method for flocculation through pH modulation. Bioresource Technology, 213, 216–221. <https://doi.org/10.1016/J.BIORTECH.2016.03.114>
- Unterlander, N., Champagne, P., & Plaxton, W. C. (2017). Lyophilization pretreatment facilitates extraction of soluble proteins and active enzymes from the oil‐accumulating microalga Chlorella vulgaris. Algal Research, 25, 439–444. <https://doi.org/10.1016/j.algal.2017.06.010>
- Vandamme, D., Foubert, I., Fraeye, I., Meesschaert, B., & Muylaert, K. (2012). Flocculation of Chlorella vulgaris induced by high pH: Role of magnesium and calcium and practical implications. Bioresource Technology, 105, 114–119. [https://doi.org/10.1016/j.biortech.](https://doi.org/10.1016/j.biortech.2011.11.105) [2011.11.105](https://doi.org/10.1016/j.biortech.2011.11.105)
- Vochozka, M., Horák, J., Krulický, T., & Pardal, P. (2020). Predicting future brent oil price on global markets. Acta Montanistica Slovaca, 25(3), 375–392. <https://doi.org/10.46544/AMS.v25i3.10>
- Vochozka, M., Rowland, Z., Suler, P., & Marousek, J. (2020). The influence of the international price of oil on the value of the EUR/USD exchange rate. Journal of Competitiveness, 12(2), 167–190. [https://](https://doi.org/10.7441/joc.2020.02.10) doi.org/10.7441/joc.2020.02.10

- Wan, C., Alam, M. A., Zhao, X. Q., Zhang, X. Y., Guo, S. L., Ho, S. H., Chang, J. S., & Bai, F. W. (2015). Current progress and future prospect of microalgal biomass harvest using various flocculation technologies. Bioresource Technology, 184, 251–257. [https://doi.org/](https://doi.org/10.1016/j.biortech.2014.11.081) [10.1016/j.biortech.2014.11.081](https://doi.org/10.1016/j.biortech.2014.11.081)
- Wang, J., & Yin, Y. (2018). Fermentative hydrogen production using pretreated microalgal biomass as feedstock. Microbial Cell Factories, 17(1), 22. <https://doi.org/10.1186/s12934-018-0871-5>
- Wang, S. K., Stiles, A. R., Guo, C., & Liu, C. Z. (2015). Harvesting microalgae by magnetic separation: A review. Algal Research, 9, 178–185. <https://doi.org/10.1016/j.algal.2015.03.005>
- Wei, C., Huang, Y., Liao, Q., Zhu, X., Xia, A., & Zhu, X. (2020). Application of bubble carrying to Chlorella vulgaris flocculation with branched cationic starch: An efficient and economical harvesting method for biofuel production. Energy Conversion and Management, 213, 112833. <https://doi.org/10.1016/j.enconman.2020.112833>
- Wittkamp, F., Senger, M., Stripp, S. T., & Apfel, U. P. (2018). [FeFe]-Hydrogenases: Recent developments and future perspectives. Chemical Communications, 54(47), 5934–5942. [https://doi.org/10.](https://doi.org/10.1039/c8cc01275j) [1039/c8cc01275j](https://doi.org/10.1039/c8cc01275j)
- Wojtyła, S., Klama, P., & Baran, T. (2017). Is 3D printing safe? Analysis of the thermal treatment of thermoplastics: ABS, PLA, PET, and nylon. Journal of Occupational and Environmental Hygiene, 14(6), D80–D85. <https://doi.org/10.1080/15459624.2017.1285489>
- Xie, D., Ji, X., Zhou, Y., Dai, J., He, Y., Sun, H., Guo, Z., Yang, Y., Zheng, X., & Chen, B. (2022). Chlorella vulgaris cultivation in pilot‐scale to treat real swine wastewater and mitigate carbon dioxide for sustainable biodiesel production by direct enzymatic transesterification. Bioresource Technology, 349, 126886. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.biortech.2022.126886) [biortech.2022.126886](https://doi.org/10.1016/j.biortech.2022.126886)
- Yang, B., Liu, J., Ma, X., Guo, B., Liu, B., Wu, T., Jiang, Y., & Chen, F. (2017). Genetic engineering of the Calvin cycle toward enhanced photosynthetic CO2 fixation in microalgae. Biotechnology for Biofuels, 10(1), 229. <https://doi.org/10.1186/s13068-017-0916-8>
- Yin, Z., Zhu, L., Li, S., Hu, T., Chu, R., Mo, F., Hu, D., Liu, C., & Li, B. (2020). A comprehensive review on cultivation and harvesting of microalgae for biodiesel production: Environmental pollution control and future directions. Bioresource Technology, 301, 122804. [https://doi.org/10.](https://doi.org/10.1016/j.biortech.2020.122804) [1016/j.biortech.2020.122804](https://doi.org/10.1016/j.biortech.2020.122804)
- Yoksan, R., Boontanimitr, A., Klompong, N., & Phothongsurakun, T. (2022). Poly(lactic acid)/thermoplastic cassava starch blends filled with duckweed biomass. International Journal of Biological Macromolecules, 203, 369–378. <https://doi.org/10.1016/j.ijbiomac.2022.01.159>
- Yusof, Y. A. M., Md. Saad, S., Makpol, S., Shamaan, N. A., & Ngah, W. Z. W. (2010). Hot water extract of Chlorella vulgaris induced DNA damage and apoptosis. Clinics, 65(12), 1371–1377. [https://doi.org/10.1590/](https://doi.org/10.1590/S1807-59322010001200023) [S1807-59322010001200023](https://doi.org/10.1590/S1807-59322010001200023)
- Zabochnicka, M., Krzywonos, M., Romanowska‐Duda, Z., Szufa, S., Darkalt, A., & Mubashar, M. (2022). Algal biomass utilization toward circular economy. Life, 12(10), 1480. [https://doi.org/10.3390/](https://doi.org/10.3390/life12101480) [life12101480](https://doi.org/10.3390/life12101480)
- Zeller, M. A., Hunt, R., Jones, A., & Sharma, S. (2013). Bioplastics and their thermoplastic blends from Spirulina and Chlorella microalgae. Journal of Applied Polymer Science, 130(5), 3263–3275. [https://doi.org/10.](https://doi.org/10.1002/app.39559) [1002/app.39559](https://doi.org/10.1002/app.39559)
- Zhu, L. D., Hiltunen, E., & Li, Z. (2019). Using magnetic materials to harvest microalgal biomass: Evaluation of harvesting and detachment efficiency. Environmental Technology, 40(8), 1006–1012. [https://](https://doi.org/10.1080/09593330.2017.1415379) doi.org/10.1080/09593330.2017.1415379

How to cite this article: Al‐Hammadi, M., & Güngörmüşler, M. (2024). New insights into Chlorella vulgaris applications. Biotechnology and Bioengineering, 121, 1486–1502. <https://doi.org/10.1002/bit.28666>